

**Effective from AY: 2022-23**

<b><u>Prerequisites for the course:</u></b>	Basic knowledge of microbial habitats in a tropical setup and general idea of diseases affecting crops.	
<b><u>Objective(s):</u></b>	To impart requisite field and lab skills in plant microbiology and pathology with emphasis on tropical strains and local needs in agriculture and economy dealing with economically important microbes.	
<b><u>Content:</u></b>	<p><b>Microbiology</b></p> <ol style="list-style-type: none"> <li>1. Microbial ecology in relation to the plants-Introduction to field techniques to study plant-microbe interactions. <b>2 hours</b></li> <li>2. Isolation of Phylloplane microflora on microbiological media and visualization of colony characteristics. <b>4 hours</b></li> <li>3. Isolation of Rhizosphere microflora on microbiological media and visualization of colony characteristics. <b>4 hours</b></li> <li>4. Isolation of endophytes and visualization of colony characteristics. <b>4 hours</b></li> <li>5. Maintenance of pure cultures of phylloplane, Rhizosphere and endophytic microflora using common microbiological media. <b>2 hours</b></li> <li>6. Use of Microscopy in studying microbes in detail - preparation of unstained and stained specimens of eubacteria, actinobacteria. Photomicrography and digital image analysis of representative pure cultures and interpretation of results. <b>2 hours</b></li> <li>7. Preparation of unstained and stained specimens of yeasts, fungi. Examination of gram character of bacteria. <b>2 hours</b></li> <li>8. SEM study of bacteria, fungi, plant viruses using electron dense stains. <b>2 hours</b></li> <li>9. Studying Phylogeny of plant viruses using bioinformatics tools. <b>2 hours</b></li> <li>10. Study of root nodulation, symbiosome, <i>Rhizobium</i>, leghemoglobin and Quorum Sensing in bacterial population. <b>2 hours</b></li> <li>11. Methods of isolation and culturing of fungi: colony characters; microscopic observations; morphology of hyphae and spores. <b>2 hours</b></li> <li>12. Study of reproductive structures of different genera of fungi. <b>2 hours</b></li> <li>13. Study of fungal physiology in pure colonies – characterization of fungal colonies. <b>2 hours</b></li> <li>14. Microfluidics in mycology- fabrication and application of microfluidics devices to fungal cultures for real time visualization of fungal metabolic activities. <b>2 hours</b></li> <li>15. Introduction to mycological databases and myco-systematics on Internet. <b>2 hours</b></li> <li>16. Introduction to Mycobioinformatics- tools and techniques (exercise to construct fungal phylogenetic tree to be given). <b>2 hours</b></li> </ol>	

	<p>17. Observation of different fungal substrates using sterile moist chamber incubation (e.g. herbivore dung; decomposing leaf-litter). <b>2 hours</b></p> <p>18. Observations on ecological succession of fungi; Terrestrial, marine and freshwater fungi. <b>2 hours</b></p> <p>19. Particle-plating technique for isolation of litter fungi. <b>2 hours</b></p> <p>20. Technique for isolation of fungal endophytes. <b>2 hours</b></p> <p>21. Isolation and serial dilution techniques (e.g. soil, dung and leaf litter). <b>2 hours</b></p> <p><b>Plant pathology</b></p> <p>22. Collection of infected specimens in the field and observation of symptoms. <b>2 hours</b></p> <p>23. Hand sections and tease mounts from infected plant specimens. <b>2 hours</b></p> <p>24. Study of viral, bacterial and fungal diseases of crop plants (cereal, vegetable, fruit, and plantations) from surrounding habitats in Goa. <b>4 hours</b></p> <p>25. Submission of 10 dried herbarium specimens of infected plant materials [fungal (4) +bacterial (3) + viral (3)] collected from nearby habitats. <b>2 hours</b></p> <p>26. A mini field project to study crop diseases from field and market specimens. <b>4 hours</b></p> <p><i>All plant pathology practicals will be conducted and any 16 hours from microbiology component will be conducted depending on availability of material, chemicals, equipments, etc.</i></p>	
<b><u>Pedagogy:</u></b>	Field visits and lab exercises/sample collections/use of electronic, digital and visual keys, herbarium production/videos/moodle guided exercises/mini projects/demonstration.	
<b><u>References/ Readings:</u></b>	<p><b>Agrios, G.N.</b> (1997). Plant Pathology. Academic Press, New Delhi.</p> <p><b>Bilgrami, K.S. and Dube, H. C.</b> (1990). A text book of Modern Plant Pathology. Vikas Publishing House, New Delhi.</p> <p><b>Butler, E.J. and Jones, S. G.</b> (1949). Plant Pathology. Mc Millan, London.</p> <p><b>Chatterjee, P.B.</b> (1997). Plant Protection Techniques. Bharati Bhavan, Patna.</p> <p><b>Chattopadhyay, S.B.</b> (1991). Principles and Procedures of Plant Protection. Oxford &amp; IBH, New Delhi.</p> <p><b>Sharma, P.D.</b> (2004). The Fungi for University students. Rastogi Publications, Meerut.</p> <p><b>Srivastava, J.P.</b> (1998). Introduction to Fungi. Central Book Depot, Allahabad.</p> <p><b>Sumbali, G.</b> (2005). The Fungi. Narosa Publishing House, New Delhi.</p>	

<b><u>Learning Outcomes:</u></b>	<ol style="list-style-type: none"> <li>1. Ability to work as a field microbiologist to sample various habitats and asplant pathologist being able to identify disease symptoms.</li> <li>2. Being able to identify common micro and macrofungi from diverse natural habitats.</li> <li>3. Being able to prepare herbarium of diseased plants.</li> <li>4. Being able to isolate and manage microbial cultures.</li> <li>5. Being able to perform image analysis of cultures.</li> <li>6. Being able to apply techniques learnt in appropriate projects involving economically important microbes.</li> </ol>	
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