Name of the Programme: M.Sc. Part-I (Biochemistry)

Course Code: CHB-503 Title of the Course: Cell and Developmental Biology

Number of Credits: 4

Effective from AY: 2022-23

Pre-requisites	Students should have graduate level knowledge either in ch	emical or life
for the Course:	sciences or should have qualified change of discipline test.	
Course Objectives:	<ol> <li>The objective is to offer detailed knowledge about cell bic cellular organelles, the communication pathways associated processes.</li> <li>Introduction of the fundamental concepts of organismal obiology.</li> <li>The course aims to provide the students insights on bas techniques and their current applications.</li> </ol>	d with cellular developmental
Content:		No of hours
	<ol> <li>Structural organization of the cell         <ul> <li>Prokaryotic and eukaryotic cells.</li> <li>Animal and plant cells.</li> <li>Structure and functions of cellular and subcellular organelles.</li> </ul> </li> </ol>	
	<ul> <li>2. Biological membrane structure and function         <ul> <li>a. Structure and functions of membrane.</li> <li>b. Transport across cell membrane.</li> <li>c. Passive and active transport of molecules across biological membranes.</li> <li>d. membrane pumps.</li> </ul> </li> </ul>	5
	<ul> <li>3. Cell division and cell cycle <ul> <li>a. Mitosis.</li> <li>b. Meiosis.</li> <li>c. Regulation of the cell cycle.</li> </ul> </li> </ul>	5
	<ul> <li>4. Cellular communication and Cell signalling <ul> <li>a. Signal transduction pathway.</li> <li>b. Signalling molecules and their receptors.</li> <li>c. G-Protein Coupled receptors.</li> <li>d. Receptor Tyrosine Kinases.</li> <li>e. MAP kinase pathway and JAK-STAT pathway.</li> <li>f. Light signalling in plants.</li> <li>g. Bacterial chemotaxis and quorum sensing.</li> <li>h. Programmed cell death (Apoptosis).</li> </ul> </li> </ul>	10
	<ul> <li>5. Fundamentals of organismal development <ul> <li>a. Potency, commitment, specification, induction,</li> <li>competence.</li> <li>b. Determination and differentiation, morphogenetic</li> <li>gradients.</li> <li>c. Cell fate and cell lineages.</li> <li>d. Stem cells, genomic equivalence.</li> <li>e. Cytoplasmic determinants, imprinting and mutants.</li> </ul> </li> </ul>	6

	6. Early organismal development	6
	a. Gametogenesis.	_
	b. Cell surface molecules in sperm-egg recognition in	
	animals.	
	c. Embryo sac development and double fertilization in	
	plants.	
	d. Zygote formation, cleavage, blastula formation,	
	embryonic fields gastrulation.	
	e. Formation of germ layers in animals, embryogenesis.	
	f. Establishment of symmetry in plants.	
	g. Seed formation.	
	7. Plant tissue culture: techniques and applications	6
	a. Introduction to plant tissue culture and various	
	requirements.	
	b. Preparation for tissue culture.	
	i. Surface sterilization of plant tissue material.	
	ii. Basic procedure for aseptic tissue transfer.	
	c. Tissue culture methodologies.	
	i. Callus Culture.	
	ii. Cell Suspension Culture, protoplast culture and	
	hybridization.	
	iii. Organogenesis.	
	iv. Plant micropropagation.	
	v. Somatic Embryogenesis.	
	vi. Incubation and maintenance of culture.	
	d. Applications of PTC.	
	8. Animal tissue culture: techniques and applications	6
	a. Introduction to animal tissue culture and various	
	requirements.	
	b. Typical cell lines, growing mammalian cells and general	
	maintenance of cells.	
	c. Applications of ATC.	
	9. Microbial culture techniques	6
	a. In vitro culture techniques.	
	b. Nutrient requirements.	
	c. Applications in industry.	
Pedagogy:	Mainly lectures and tutorials. Seminars / term papers /	assignments /
	presentations / self-study or a combination of some of these ca	n also be used.
	ICT mode should be preferred. Sessions should be interactive in na	ature to enable
	peer group learning.	
References/	1. Karp, G.; Cell and Molecular Biology: Concepts and experimen	ts; John Wiley
Readings:	and Sons Inc., 2015; 8 <sup>th</sup> Edition.	,,
	2. Lodish, H.; Berk A.; Kaiser, C. A; Krieger, M.; Bretscher, A.;	HiddePloegh.
	Amon A.; Martin, K. C.; Molecular Cell Biology; W.H. Freeman	_
	2016; $8^{\text{th}}$ Edition.	
	3. Freshney, I.; Culture of Animal Cells: A Manual of Basic T	echnique and
	Specialized Applications; Wiley-Blackwell; 2016; 7 <sup>th</sup> Edition.	
		cular Biology
	4. DeRobertis, E.D.P.; DeRobertis Jr. E.M.F; Cell and Molection	cului Diology, I

	<ol> <li>5. Pelczar, M.; Reid, R.D.; Chan E.C.S.; Microbiology. MacGraw-Hill; 2001; 5<sup>th</sup> Edition.</li> <li>6. Smith, R.H.; Plant tissue culture: technique and experiments; Academic Press; 2012; 3<sup>rd</sup> Edition.</li> </ol>
	<ol> <li>Gilbert, S.F.; Barresi M. J.; Developmental Biology; Oxford University Press; 2020; 12<sup>th</sup> Edition.</li> </ol>
Course Outcomes:	<ol> <li>Students will be able to describe the cell structure, cell division and cell cycle mechanisms, various cellular organelles and their functions.</li> <li>Students will be able to explain the processes of transport across cell membranes, various cellular communication pathways along with their significance and understand the fundamentals of developmental biology.</li> <li>The students will be able to apply the basic cell culture techniques needed to work in a biological research laboratory.</li> <li>The students will be prepared for advanced courses in life science such as Cancer biology, Neurochemistry, etc.</li> </ol>