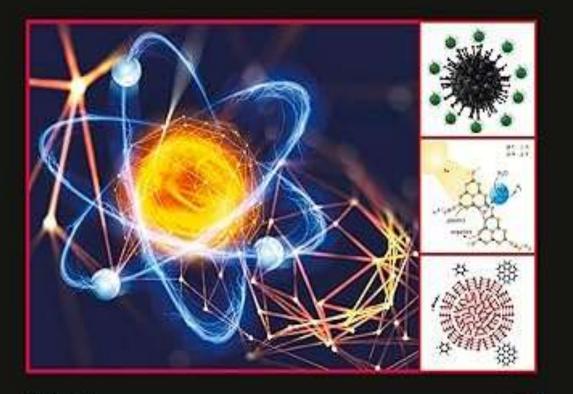
Progress in Biochemistry and Biotechnology

ADVANCES IN NANO AND BIOCHEMISTRY

Environmental and Biomedical Applications



Edited by Pranay Pradeep Morajkar Milind Mohan Naik



No. of pages: 618 Edition: 1 Imprint: Academic Press

eBook ISBN: 9780323952545

Language: English Published: May 12, 2023 Paperback ISBN: 9780323952538

Table of contents

Cover image Title page Table of Contents Copyright Dedication and acknowledgment Contributors Preface SECTION I. Environmental Studies Chapter 1. Coupling of photocatalytic and bioremediation processes for enhanced mitigation of xenobiotic pollutants from wastewater 1.1. Introduction 1.2. Xenobiotic remediation methods

- 1.3. Challenges and future outlook
- 1.4. Summary and conclusion

Chapter 2. Bioinspired nanomaterials for remediation of toxic metal ions from wastewater

- 2.1. Introduction
- 2.2. Strategies for the bioinspired nanomaterials synthesis
- 2.3. Heavy metals removal technologies employing bioinspired nanoparticles
- 2.4. Conclusions and future prospect

Chapter 3. Biocompatible nanomaterials for sensing and remediation of nitrites and fluorides from polluted water

- 3.1. Introduction
- 3.2. Nitrites and fluorides in water and wastewater
- 3.3. Preparation techniques of bionanomaterials
- 3.4. Bionanomaterials as ionic sensors
- 3.5. Nitrites and fluorides remediation by bionanomaterials
- 3.6. Challenges and future perspectives
- 3.7. Conclusions

Chapter 4. Role of gum nanostructured hydrogels in water purification, desalination, and atmospheric water harvesting applications: Advances, current challenges, and future prospective

- 4.1. Introduction
- 4.2. Fundamental principles of techniques and instrumentation procedures
- 4.3. Latest research and development in the field
- 4.4. Summary and conclusion
- 4.5. Challenges and future outlook
- Chapter 5. Versatile nanomaterials for remediation of microplastics from the environment
- 5.1. What are microplastics?
- 5.2. Effect of microplastics on human health
- 5.3. Traditional methods of microplastics separation
- 5.4. Advanced methods of microplastics separation
- 5.5. Nanomaterials
- 5.6. Remediation of microplastics using various nanomaterials
- 5.7. MPs adsorption strategies using nanomaterials
- 5.8. MPs degradation using nanomaterials

5.9. Limitations, challenges, and future outlook

Chapter 6. Plastic degradation—contemporary enzymes versus nanozymes-based technologies

- 6.1. Introduction
- 6.2. Major natural enzymes for plastic degradation
- 6.3. Major polymers that form plastic and their degradation
- 6.4. Nanozymes
- 6.5. Computational advancement for enzyme identification
- 6.6. Conclusion and future perspectives

Chapter 7. Current trends in sensing and remediation of gaseous pollutants in the atmosphere

7.1. Introduction to the gas phase chemistry and pollutants of the atmosphere (tropospheric emphasis)

7.2. Current trends in measurement approaches of important gaseous pollutants of the atmosphere and the associated challenges

- 7.3. Current trends in concentration levels and mitigation approaches
- 7.4. Challenges and future outlook

Chapter 8. Emerging nonnoble metal nanocatalysts for complete mitigation of combustion generated CO, NOx, and unburnt hydrocarbons

- 8.1. Introduction
- 8.2. Different catalytic methods for the mitigation of pollutants emission
- 8.3. Latest research and development in mitigation of pollutants emission
- 8.4. Conclusion and future outlook

Chapter 9. Advanced methodologies for remediation of combustion-generated particulate matter (soot) from the environment

- 9.1. Introduction
- 9.2. Genesis of soot
- 9.3. Latest research and development in the remediation of combustion generated soot
- 9.4. Summary and conclusion
- 9.5. Challenges and future outlook

Chapter 10. Recent advances in quantification and remediation technologies for toxic PAH mitigation from the environment

- 10.1. Introduction
- 10.2. PAH detection and quantification technologies
- 10.3. Techniques for the environmental remediation of PAHs
- 10.4. Summary and future outlook
- SECTION II. Biomedical Studies

Chapter 11. Application of nanoparticles as quorum quenching agent against bacterial human pathogens: a prospective therapeutic nanoweapon

- 11.1. General introduction
- 11.2. Nanoparticles: fundamentals and principles
- 11.3. Latest research on nanoparticles as quorum quenching agents
- 11.4. Mechanisms of quorum quenching by nanoparticles
- 11.5. Techniques and biosensors involved in quorum quenching research of nanoparticles
- 11.6. Summary and conclusion

11.7. Challenges and future prospects

Chapter 12. Biocompatible green-synthesized nanomaterials for therapeutic applications

12.1. Introduction

12.2. Fundamental principles of techniques and instrumentation/methods/procedures involved

12.3. Latest research and development in the field

12.4. Summary and conclusion

12.5. Challenges and future outlook

Chapter 13. Toxicological aspects of nanomaterials in biomedical research

13.1. Introduction

13.2. Toxicity of nanomaterials in biomedicine

13.3. Genotoxic biomarkers

13.4. Safety against toxic effects

13.5. Summary and conclusions

13.6. Challenges and future outlook

Chapter 14. Quantum dots and hybrid structures as an innovative solution for bioimaging and diagnosis of viral infections

14.1. Introduction

14.2. Synthesis methods, modification strategies, and properties of QDs

14.3. QDs photoluminescence—principles/mechanisms

14.4. Application of QDs in bioimaging and detection of viruses

14.5. Challenges and future prospects

14.5. Challenges and future prospects

14.6. Summary and conclusion

Chapter 15. Magnetic nanomaterials and their hybrids for magnetic hyperthermia

15.1. Introduction

15.2. Nanomagnetism

15.3. Magnetic alloy nanoparticles for MHT

15.4. Ferrite magnetic nanoparticles for magnetic hyperthermia

15.5. Superparamagnetic materials for magnetic hyperthermia

15.6. Summary and conclusion

15.7. Challenges and future outlook

Chapter 16. Advanced functionalized nanomaterial-based electrochemical biosensors for disease diagnosis

16.1. Introduction

16.2. Fundamental techniques of biosensing and nanomaterial-based diagnostic tools

16.3. Latest research and development in the biosensing field

16.4. Conclusion and future perspectives

Chapter 17. Recent advances in MOFs-based nanocomposites for treatment of retinopathy or retinarelated biomedical applications

17.1. Introduction

17.2. Traditional methods of drug loading for ocular disease treatment

17.3. Categories of nanocarriers

17.4. Drug delivery routes for nanocarriers

CHAPTER 13

Toxicological aspects of nanomaterials in biomedical research

Avelyno H. D'Costa, PhD¹, Shamshad Shaikh, PhD¹, Gandhita Kundaikar, MSc¹ and Swizzle Furtado, MSc²

¹School of Biological Sciences and Biotechnology, Goa University, Taleigao, Goa, India;
²Department of Zoology, Carmel College for Women, Nuvem, Goa, India

13.1 Introduction

Modern advances in nanoscience and nanotechnology have augmented the increase and production of many new nanomaterials with unique properties for industrial and biomedical uses in recent years. The extremely small size of these materials allows them to be used in a number of applications such as in drug delivery, cancer therapy, prosthetics, dentistry, pharmaceuticals, etc. With advancements in the field of medicine, a number of nanomaterials are synthesized with potential medical applications. For instance, quantum dots are used as fluorescent labels in drug delivery systems and in disease detection [1-3]. Carbon nanotubes can also be used in drug delivery systems due to their capacity to adsorb pharmaceutical formulations as well as in tissue engineering [4-6]. Liposomes, composed of lipid bilayers, are also used as excellent drug delivery carriers with the drug being adsorbed on the surface or sequestered within the core [7,8]. Magnetic nanoparticles also have a number of applications such as hyperthermia treatment, magnetic resonance imaging, and drug delivery [9-11].

However, some of these nanomaterials may exert toxic side effects on nontarget cells and tissues. They may also accumulate in various nontarget organs and exert toxic effects (Fig. 13.1). The size of these nanoparticles with high surface area and uncommon surface chemistry and reactivity can lead to unique problems for biological animals or cells and the environment. Such undesirable effects can be minimized by synthesizing biocompatible nanomaterials or surface coating or surface modifying the nanomaterial before being administered. Various strategies exist to make the nanoparticles biocompatible such as coating them with polymers like polyethylene glycol (PEG) or other biological substances such as aptamers. Nanoparticles may also be synthesized to be biodegradable such that they can be rapidly cleared from the body after therapy. However, even these formulations need to be tested prior to administration to understand possible toxicity and interaction with biomolecules in vivo and in vitro.

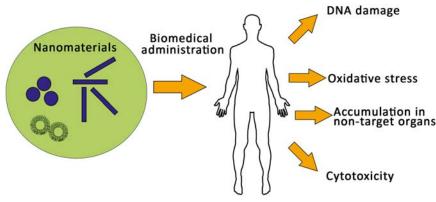


Figure 13.1 Toxic effects of nanoparticles in living organisms.

13.2 Toxicity of nanomaterials in biomedicine

13.2.1 Quantum dots

Quantum dots (QDs) are fluorescence-type semiconductor nano-sized particles used for their unique stability against photobleaching, and excitability for multicolor emission with a single light source [12], made up of either heavy metals or inorganic materials with a size ranging from 2 to 10 nm. QDs contain atom-like discrete electronic energy levels, and are therefore called artificial atoms. They consist of two free functional groups for binding with the molecules of a potential drug. Surface modification of these quantum dots through covalent and/or noncovalent binding of various substances affects and alters the properties of the drug molecules. The outer shell of QDs is made of semiconductor material that provides the surface for bio-conjugation thereby improving its aqueous solubility. This also provides an effective surface area for binding the drugs to the target molecule. Various functionalizations and surface modifications make them suitable for application in the pharmaceutical field such as biomedical imaging, drug delivery, drug release study, and diagnosis [2]. The regulatory status of quantum dots is not yet clear; but is, however, regarded as safe for usage. The first clinical trial of quantum dot technology in humans was approved by USFDA in the year 2011. With upcoming advances in technology, most of the chemotherapeutic and cytotoxic drugs are delivered via quantum dots for improved pharmacological action [2].

Owing to their wide application in the medical field, knowledge of the toxicity of QDs becomes imperative. The toxicity of quantum dots depends upon the size, dose, material used, route of exposure or administration, capping material, and composition. For example, nanosized cadmium QDs are found to be highly toxic because they can easily enter tissues and cells and exert their damaging effects [13,14]. Furthermore, Cd-QDs are extremely toxic compared to non-cadmium QDs [15]. CdTe QDs have also been reported to induce toxicity in the liver and kidney which may be reversible

over a period of time [16]. In distribution and accumulation studies, QDs could be detected in various tissues of mice even after 2 years of exposure [17]. Ref. [18] reported the accumulation of QDs in the immune organs namely, the liver, spleen, lungs, and kidney after intraperitoneal injection of QDs in mice and subsequent inflammation in some of the tissues. Certain studies have revealed that QDs also exert toxic effects on the respiratory system. For instance, Ref. [19] studied the in vivo effects of novel graphene QDs with potential applications in biomedicine. They reported fibrosis evidenced by increase of collagen I and expression of profibrotic genes TGF- β 1 and p-Smad3 in the lungs of mice exposed intranasally to QDs. In another study, black phosphorus quantum dots which have been used extensively in biomedicine were found to induce significant cytotoxicity and decreased cell viability in human-derived lung cells in vitro probably brought about by ROS [20]. Lung inflammation and injury are also reported in several QD exposure studies [21–23]. Surface modification thus becomes very important to reduce toxicity and simultaneously impart biocompatibility, stability, and specificity to QDs [15].

13.2.2 Carbon nanoparticles

Carbon nanoparticles are used extensively in biomedical applications due to their excellent biocompatibility and relatively low toxicity [24]. Carbon nanoallotropes like fullerenes, carbon dots, graphene sheets, graphene oxide, carbon nanotubes, and nano-diamonds have been used in the medical field with applications in pharmacy and medicine for drug delivery system in therapeutics, cancer therapy, regenerative studies [25-28]. In spite of these applications, some studies have reported the toxicity of carbon nanoparticles. Ref. [29] reported that multiwalled carbon nanotubes induced various degrees of cytotoxicity in human macrophages in vitro. Similarly Ref. [30] observed cytotoxicity in HeLa cells exposed to carbon nanoparticles. They further reported that cytotoxicity could be attenuated by adsorbing serum proteins onto the nanoparticles. Carbon nanotubes have been reported to affect the epithelial cells of lungs in a number of studies. For instance, Ref. [31] compared the toxicity of multiwalled carbon nanotubes and onion-like shellshaped carbon nanoparticles in human bronchial epithelial cells and found that the carbon nanotubes were more toxic. Similarly Ref. [32] reported that carbon nanotubes induced higher cytotoxicity than carbon nanoparticles in human epithelial cell line A549. Singlewalled carbon nanotubes were also found to induce fibrosis in the lungs of mice as well as elevated production of inflammatory cytokines [33]. The essential mechanisms by which carbon nanotubes induce toxicity include oxidative stress by generating ROS, inflammatory responses, malignant transformation, DNA damage and mutation (errors in chromosome number as well as disruption of the mitotic spindle), the formation of granulomas, and interstitial fibrosis [34].

Other carbon nanoparticles such as fullerenes (C60) also have applications in drug and gene delivery as well as in dermatology [35,36]. However, their toxicity has been reported by several researchers in lungs [37-39] (Marchione et al., 2007). The possible mechanisms of toxicity of fullerenes in lung cells include induction of ROS and DNA damage [40].

13.2.3 Metallic nanoparticles

Metallic nanoparticles have a number of biomedical applications depending on the type of metal used. The most commonly employed metals used in biomedical applications are gold, silver, zinc, iron, and manganese and have been extensively studied with regard to their toxicity and biocompatibility. A number of these nanoparticles are regularly used as drug carrier molecules in the treatment of neoplastic malignancies [41]. Metallic NPs allow effective conveyance of anticancer drugs into the tumors by utilizing the pathophysiology of the neoplastic cells, thereby modifying the therapeutic outcomes [42]. These metallic nanoparticles are used by targeting ligands with specific receptors on tumor cells thus offering targeted delivery to the tumor cells thereby evading the nonneoplastic cells [43,44]. Metallic nanoparticles also accumulate in various cells such as macrophages and other immune cells and various strategies are adopted to avoid uptake by these cells for effective drug delivery [45-47]. The size, density, surface charge, and chemistry of metal NPs play a crucial role in generating an immune response and renal clearance, thus increasing the circulation time and bioavailability for the target organs [48–50]. Once administered, they accumulate in immune cells, detoxifying and excretory tissues such as the liver, spleen, lymph nodes, bone marrow, adrenal glands, and kidneys [51-54].

In the recent years, gold nanoparticles are receiving much recognition and attention in medical research due to its unusual physical properties such as high resistance to corrosion which makes it biocompatible for medical applications like diagnosis and treatment of several diseases such as cancer, skin ulcers HIV, smallpox, measles, and rheumatoid arthritis and also has possible promising applications as an anticancer, antibacterial, and bio-diagnostic material [55–57]. Gold nanoparticles are also valued in the medical field due to their relative chemical stability that makes the preparation and fabrication methods modest, straightforward, and less dangerous. Gold nanoparticles are known to be less toxic compared to other metal nanoparticles (although not completely nontoxic), this is the reason they are more appropriate for biological/medical applications. Some studies have reported the toxicity of gold nanoparticles. Ref. [58] injected various sizes of gold nanoparticles in mice and observed sickness and altered pathology of organs in mice injected with gold nanoparticles with a size range of 8–37 nm. The size, concentration, and exposure conditions of gold nanoparticles have been reported to influence their cytotoxicity. For instance, gold nanoparticles were observed to enter human dermal fibroblast cells via clathrin-mediated endocytosis and accumulate in vacuoles where they can exert cytotoxicity [59]. Surface charge, whether charged or neutral, may also be an important factor for the mechanism of toxicity of gold nanoparticles [60].

Silver nanoparticles, another widely used metal nanoparticle, are present in several personal care products owing to their excellent antimicrobial activities. However, several studies have concluded that they are toxic both in vivo and in vitro which may be dependent on their size, shape, surface coating, and surface charge [61–63]. Ref. [64] orally gavaged rats with silver nanoparticles and observed several toxic effects such as decrease in body weight, alterations of alkaline phosphatase and cholesterol, and liver damage. Silver nanoparticles were found to be retained in the brain and testes of rats even 8 weeks after a 28-day exposure which could lead to probable long-term toxic effects [65]. In another study, silver nanoparticles were found to leach silver ions when dispersed in water which was found to influence toxicity in murine macrophages [66]. The probable mechanisms of silver nanoparticle toxicity include generation of ROS, interaction with cell membranes and subsequent damage, protein and nucleic acid interaction triggering various cell signaling pathways [67]. Silver nanoparticles have also been reported to impair mitochondrial function such as uncoupling of oxidative phosphorylation [68].

Zinc oxide is generally recognized as safe (GRAS) by the FDA [69], but at nanoscale level, they are known to attain novel properties which are not present in the micro or larger sizes. Studies carried out in mice reveal the potential of ZnO nanoparticles to accumulate in organs like the liver, spleen, heart, bone, and the pancreas [70]. The toxicity of ZnO NPs on mammals has been demonstrated both in vitro and in vivo. Inhalation of these nanoparticles are known to cause potent yet reversible pulmonary inflammation [71]. Intratracheal delivery of ZnO NPs into the lungs of rats induced eosinophilia, propagation of airway epithelial cells, goblet cell hyperplasia, and pulmonary fibrosis [72]. It is believed that the mechanism of ZnO nanoparticle toxicity is through production of ROS [69] and dissolution of lysosomal membranes [72]. A three-tier model of ROS oxidative stress is described by Ref. [73]. Tier one includes increases in antioxidant enzymes to start the initial antioxidant defense, followed by Tier two which includes an increase in potent proinflammatory cytokines leading to inflammation, while Tier three is characterized by mitochondrial perturbation resulting in cellular death by apoptosis or necrosis. All three tiers have been observed for ZnO nanoparticles in immortalized phagocytic or bronchial epithelial cells leading to damage of lipids, proteins, and DNA, increased release of lactate dehydrogenase, and death by either necrosis or apoptosis [21,73-77].

Several in vivo and in vitro studies have been carried out to study the toxicity of manganese oxide (MnO) nanoparticles. Manganese as an important component of the antioxidant enzymes namely superoxide dismutase but literature has cited that manganese compounds among micronutrients cause neurotransmitter anomalies, elevate ROS level, and promote protein misfolding and aggregation, and the mechanisms contributing to the Parkinson's disease model [78–81]. MnO nanoparticles were reported to induce moderate toxicity as seen from mitochondrial reduction activity in PC-12 cells from PC-12 cell line cultured neuronal phenotype [82]. Studies have shown that MnO nanoparticles affect the reproductive system of rats having chronic exposure, decrease in the number of sperms, spermatogonia, spermatocytes, diameter of seminiferous tubules, and also bring down the motility of sperms [83]. At lower doses MnO nanoparticles induce genotoxicity, biochemical, and histopathological alterations as compared to microparticles of MnO. Biodistribution investigations have revealed the presence and accumulation of MnO NPs in the gastrointestinal tract and in the organs and tissues. The concentrates of MnO NPs were also found in the liver, spleen, kidney, heart, blood, brain, and lungs. The bulk of NPs were found in the liver, kidney, as well as spleen [84].

13.2.4 Magnetic nanoparticles

Magnetic nanoparticles mainly incorporate metal nanoparticles, metal oxide nanoparticles, and metal alloy nanoparticles. Magnetic nanoparticles have a great specific surface area, which is ideal for carrying large amounts of DNA fragments, drugs and modified compounds. Additionally, most modified magnetic nanoparticles have excellent biocompatibility and superparamagnetism which make them excellent vehicles for targeted drug delivery [85]. Iron oxide nanoparticles are the most studied and commonly used magnetic nanoparticles for biomedical applications, due to their unique chemical, biological, and magnetic properties such as chemical stability, biocompatibility, high saturation magnetization, and high magnetic susceptibility. Iron oxide nanoparticles have been studied for the use in magnetic hyperthermia treatment, targeted drug delivery, and contrast agents in magnetic resonance imaging (MRI). Furthermore, a number of studies prove that magnetic nanoparticles, coated or uncoated, are not toxic which is useful for further development of their biomedical applications [86–91]. However, some magnetic nanoparticles have been proven to be toxic. The toxicity of magnetic nanoparticles depends on their size, shape, coatings, and nature of magnetically responsive components such as iron, nickel, cobalt, etc. [92]. For instance, composite iron oxide nanoparticles with silica core or nanoflakes induced significant cytotoxicity in human umbilical vein endothelial cells [91]. On the other hand, magnetic nanoparticles coated with amorphous silica were found to be protective against the toxicity of uncoated magnetic nanoparticles in human bronchial epithelium-derived BEAS-2B cells, in vitro compared to bare magnetic nanoparticles [93]. Careful administration of magnetic nanoparticles in the body is important because unintended inhalation or ingestion of these nanoparticles could possibly result in the distribution of nanoparticles in different parts of the body leading to toxicity [94–96]. The possible mechanisms of magnetic nanoparticle toxicity include oxidative stress by generation of free radicals, lipid peroxidation, and elevation of antioxidant enzymes [97-99]. This mechanism can be considered ambiguous in the sense that these nanoparticles can induce cytotoxicity and cell death in cancer cells as well as normal cells and thus careful targeting is of utmost importance [100].

13.2.5 Polymer and liposome-based nanoparticles

Polymer-based nanoparticles are colloidal carriers composed of natural polymers such as starch, polypeptides, chitin, cellulose, and polyhydroxyalkanoates (PHAs) or synthesized polymers such as polyethylene glycol (PEG), poly-lactic-acid-co-glycolic-acid (PLGA), polybutylcyanoacrylate (PBCA), and polyvinyl alcohol (PVA). They are known to effectively carry drugs, proteins, and DNA to target cells and tissues and hence synthesized as polymer nanospheres and nanocapsules. Certain disadvantages of using synthesized polymers in medical applications include nonbiodegradability and presence of toxic residues which are not properly removed during the manufacturing process [101,102]. High concentrations of polybutylcyanoacrylate nanoparticles were reported to be toxic to cell cultures but were found to be nontoxic to rats in vivo [103]. In another study, the cytotoxicity of PLGA nanoparticles coated with various stabilizers such as chitosan, PVA, and poloxamer 188 was evaluated in human-like THP-1 macrophages [104]. The PLGA nanoparticles coated with these stabilizers at high concentrations induced significant cytotoxicity while the stabilizers themselves did not induce significant cytotoxicity. One study attributed the in vitro toxicity of polymeric nanoparticles to generation of reactive oxygen species and expression of tumor necrosis factor alpha [105].

Liposome-based nanoparticles consist of cationic lipids in a bilayer that has a vesicular structure. Drugs of interest are encapsulated in the aqueous core of these liposomes for subsequent delivery to target tissues/cells. Thus, liposomes are highly biocompatible and biodegradable allowing for their extensive in biomedical applications such as targeted drug delivery [106]. Some of the disadvantages of liposome nanoparticles include low rate of encapsulation of water-soluble drugs and poor stability of the liposome. To solve the problem of stability, PEGylation of lipid nanoparticles have been used to further stabilize the nanocarriers for drug delivery [107]. However, a few studies have reported the toxicity of liposome nanoparticles. For instance, various concentrations of cationic liposomes, when injected in rats, were found to induce DNA strand breaks in the cells of the lung and spleen [108]. The expression of cytokines was also found to be elevated in the lungs, spleen, and liver of these rats. In another study, Ref. [109] reported the cytotoxicity and genotoxicity of cationic liposomes in human hepatocyte cells (HepG2) and lung epithelial cells (A549) in vitro. In an attempt to understand drug efficacy with liposomes Ref. [110] administered liposomal doxorubicin in Walker 256 tumor-bearing rats. When compared with rats administered with dendrimer doxorubicin, the liposomal formulation induced significant systemic toxicity and cardiotoxicity although both formulations displayed similar antitumor efficacy.

13.3 Genotoxic biomarkers

Genotoxicity tests are routinely used to assess the DNA damaging effects of nanoparticles before they are used for their potential applications. Two commonly used tests for assessing genotoxicity are the micronucleus test and the single cell gel electrophoresis assay (comet assay) (Fig. 13.2).

13.3.1 Micronucleus test

The micronucleus test is a rapid and easy test to assess clastogenicity (chromosomal damage) in cells. In eukaryotic cells, the chromosomes exist within a compact nucleus. However, the presence of DNA damaging agents can affect the integrity of these chromosomes such that they can cause breaks or fragmentation. Furthermore, the toxicants may also affect the spindle apparatus of cells and may not allow proper segregation of chromosomes during mitosis. This eventually leads to a broken chromosomal fragment or a whole chromosome persisting in the cytoplasm near the main nucleus, hence termed micronucleus. Thus, higher the incidence of micronuclei in cells, higher is the DNA damaging potential of the toxicant. The micronucleus test has been used as a biomarker of genotoxicity in several studies involving nanoparticle toxicity. Ref. [111]. used the in vitro cytokinesis block micronucleus assay in Chinese hamster lung fibroblast cells (V79) to test the genotoxic potential of zinc oxide nanoparticles and reported significant induction of micronuclei at higher nanoparticle concentrations. In another study, cerium

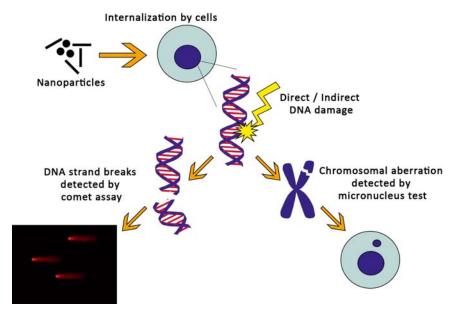


Figure 13.2 DNA damage mechanisms measured by the comet assay and micronucleus test.

oxide nanoparticles were found to induce significant micronuclei in lymphocytes in vitro as measured by the cytokinesis block micronucleus test barely 3-24 h after nanoparticle exposure [112]. The micronucleus test has also been used to report the genotoxicity of silver nanoparticles in vitro [113,114] and in vivo [51,115]. Engineered nanoparticles with potential industrial applications have also been evaluated for genotoxicity using the micronucleus test [116]. A number of studies have suggested that micronuclei formation may be attributed to oxidative stress induced by the nanoparticles [117–119].

13.3.2 Single cell gel electrophoresis assay (comet assay)

In contrast to the micronucleus test, the comet assay measures DNA damage in the form of single- or double-stranded DNA breaks. The protocol for comet assay is a bit more laborious, but is highly sensitive and allows for the visualization of DNA strand breaks in cells. In this test, cells are embedded on agarose gel-coated slides and subject to lysis. The lysis buffer degrades the membranes and cytoplasmic contents of the cell leaving only the nucleus intact. The DNA are then unwound using an alkaline or neutral unwinding buffer so as to liberate the broken DNA fragments. After a brief period of unwinding, the slides are subjected to electrophoresis where the broken DNA fragments migrate out from the nucleus under the influence of the electric field (since DNA is negatively charged). The slides are then placed in neutralization buffer and stained with a nuclear fluorescence dye such as ethidium bromide or SYBR green and observed under a fluorescence microscope with appropriate filters. Cells with a large amount of breaks show the appearance of a "comet" (the head resembling the intact DNA and tail representing broken DNA fragments which migrated out during electrophoresis). Thus, longer the tail, larger is the extent of DNA damage. Cells with no breaks resemble only the head of the comet with no tail. The tail length can be measured using visual scoring methods or with the help of software and can be recorded. DNA damage induced in this way may be due to direct effects of the nanoparticle or indirect effects mediated by free radicals. The comet assay is thus a suitable test for measuring the extent of DNA damage induced by nanomaterials [120]. Ref. [112] reported significant DNA damage in human lymphocytes in vivo measured by the alkaline comet assay. Ref. [121] also reported DNA strand breaks and DNA oxidation by various nanoparticles. They further reported that this DNA damage was brought about due to increased levels of reactive oxygen species and reduced DNA repair capacity. Similarly, the comet assay was also used to detect the genotoxicity of titanium dioxide intra-tracheally instilled in rats where they reported no genotoxicity [122]. Consequently, DNA damage measured by the comet assay was reported in human peripheral blood cultures exposed to titanium dioxide nanoparticles in vitro [123]. The comet assay was also used to detect DNA damage in human keratinocytes exposed to citrate-coated silver nanoparticles in vitro indicating the probable genotoxicity of cosmetic products [124].

13.4 Safety against toxic effects

13.4.1 Biocompatibility

Biocompatibility is a general term used to define the compatibility of biomaterial (biological compound, nanoparticle, or implantable devices) to perform its specific function in the living tissue or a living system with respect to a medical therapy without inducing any undesirable effects, that will assure minimal toxic effects, potential to cause injury, or physiological/immunological reactivity [125]. No compound/particle can be claimed to be completely biocompatible, but the research is continued to design and establish sufficiently biocompatible particles along with the benefits that it offers. To study NPs biocompatibility, there are several aspects which need to be explored, which include evaluation of cell viability, cytotoxicity, proliferation, apoptosis/necrosis, cellular morphology alteration, oxidative stress, inflammatory response, and hemotoxicity. The biocompatibility of certain nanoparticles depends closely on various several factors, which include physicochemical properties of the nanoparticles, the impact of their size, shape, and surface charge state, their formulation, biological target, dose, the methodology employed to evaluate their toxicity, and how they are delivered into the body (Huang et al., 2017) (Fig. 13.3).

One of the ways of protecting the body against the toxic effects of nanoparticles is by using a protective coating against the uncontrolled release of metal ions into the cells and tissues. A number of studies have compared the toxicity of uncoated and coated nanoparticles. For instance, Ref. [126] reported that uncoated iron oxide nanoparticles induced significant cytotoxicity in A549 and HeLa cells. However, the nanoparticles coated with a thin shell of silica reduced their toxicity rendering them more stable within biological environments. Magnetic nanoparticles used in hyperthermia treatment have also been made biocompatible with various surface modifications/coatings such as

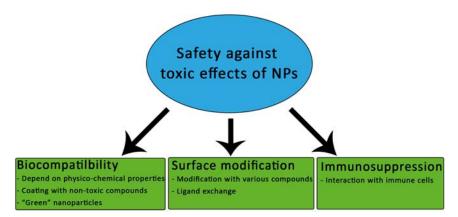


Figure 13.3 Safety against toxic effects of nanoparticles in biomedicine.

PEGylated curcumin, aptamers (small single-stranded DNA or RNA), polycations, and amino acids, to name a few [127–130].

Ref. [131] reported that gold nanoparticles loaded with doxorubicin hydrochloride (drug delivery system) were found to be cytotoxic in A549 cells. However, when they stabilized the nanoparticle—drug complex with xanthan gum, cytotoxicity was found to be significantly reduced. Similarly silver nanoparticles used in antibacterial or anti-fungal applications were stabilized with type I collagen and this composite was found to be nontoxic to fibroblasts and keratinocytes while exhibiting toxicity to bacteria [132]. In another study, zinc oxide nanoparticles, which are known to be toxic, were incorporated with nanostructured hydroxyapatite, were found to be nontoxic when administered to rats in vivo [133]. With such biocompatibility, this nanoparticle composite has potential applications in bone regeneration in orthopedics and dentistry while also acting as an antibacterial agent.

Carbon nanoparticles have been used in several biomedical applications such as bioimaging and drug carriers, but are known to induce toxicity. Thus the biocompatibility of carbon nanoparticles should be assessed before being used in biomedicine. Ref. [134] synthesized carbon dots hybridized with plasmonic nanostructures for potential applications in diagnostic bioimaging and were found to be biocompatible with Vero epithelial kidney cells in vitro. In another study, nitrogen-doped carbon nanoparticles were found to be biocompatible for bioimaging of histidine detection in cells [135]. Carbon dots have also been coupled with aspirin to be used dually as an antiinflammatory agent and fluorescent biomarker and have also found to be nontoxic in vitro [136]. Carbon nanotubes/ poly(lactic-co-glycolic acid) composites synthesized by Ref. [137] were reported to be biocompatible for bone and tissue regeneration in rats. No significant toxicity was observed probably due to the lower concentration of the carbon nanotubes. Furthermore, this biocompatibility was similar to that of poly(lactic-co-glycolic acid) administered alone.

In recent years, nanoparticles synthesized from plants have gained attention due to their eco-friendly approach and ease of production [138,139]. These "green" nanoparticles are usually preferred over engineered ones due to their biological origin and higher biocompatibility [140]. A number of studies have proven that nanoparticles synthesized from plant sources were found to be less toxic than chemically synthesized ones and thus, more biocompatible. Ref. [141] reported that silver nanoparticles synthesized from the leaf extracts of *Nigella sativa* did not induce significant cytotoxicity in mouse mesenchymal stem cells compared to chemically synthesize silver nanoparticles. Similarly Ref. [142] also reported that silver and selenium nanoparticles synthesized from various plant extracts did not induce significant toxicity in mice in vivo. In another study, gold nanoparticles synthesized from reducing gold salts by phytochemicals in tea leaves did not induce significant toxicity in cells probably due to the phytochemicals forming a coating over the gold nanoparticles [143].

13.4.2 Surface modification

Apart from the increase in biocompatibility, NPs surface modification can be a very important tool for the modulation of the body's immune response against the specific particles as well as to fix or attenuate issues related to NPs toxicity. After NPs are injected in the blood stream, they interact with a lot of specific proteins like opsonin, complement proteins, immunoglobulins, fibronectin, and apolipoproteins (protein corona). Such interactions can modify the behavior of NPs and can trigger an immune response (Barbero et al., 2017). The NP's surface modification can be exploited to promote their escape from the immune system and to increase their half-life in blood by reducing the clearance due to macrophages of the mononuclear phagocyte system (MPS). Conversely, in the next-generation vaccines, NP surface modifications can trigger the immune response toward a specific antigen (Ahmad et al., 2019; Cappellano et al., 2019; Gu et al., 2019). Furthermore, surface modification also allows nanoparticles to be taken up by cells for effective drug delivery [144].

Surface modifications usually involve ligand exchange or ligand adsorption to improve the functionalization and prevent toxicity of engineered nanoparticles [145]. Various types of nanomaterials have characteristic chemical properties and functional groups exposed on their surface to be used in the first steps of functionalization. Superparamagnetic iron oxide nanoparticles can be easily modified by using a ligand exchange strategy based on the substitution of the original surfaces with functional groups such as a diol, amine, carboxylic acid, and thiol to improve hydrophilicity and stability which would be beneficial in biomedical applications such as imaging (Korpany et al., 2016). Conversely, in another study by Ref. [146] superparamagnetic iron nanoparticles which were amine-surface-modified, though not cytotoxic, were found to alter the osteogenic and chondrogenic differentiation of human mesenchymal stem cells in vitro. Ref. [147] reported that polydopamine surface modification of CdSe quantum dots did not induce significant in vivo toxicity in rats compared to unmodified CdSe. Another nanoparticle used in drug delivery is polylactic-co-glycolic acid (PLGA), gets opsonized by the reticuloendothelial system, and thus requires surface modification using various compounds such as human serum albumin, potassium dichromate, and oxygen plasma [148-150].

13.4.3 Immunosuppressants

The immune system of humans is a complex and important series of defense mechanisms against harmful foreign substances and pathogens. Nanoparticles in biomedical research can stimulate the immune system in an undesirable or desirable way [151]. For instance, many nanoparticles induce inflammation in the body; the same property can be used for the treatment of certain diseases like cancer. On the other hand a number of nanoparticles are known to cause immunosuppression which can be used in the treatment of inflammatory disorders or autoimmune diseases. A number of mechanisms exist on how

nanoparticles achieve immunosuppression such as interaction with macrophages, antigen presenting cells as well as B and T cells [152]. However, immunosuppression by nano-particles can also render the body susceptible to various infections that ultimately cannot be combatted by the immune system [153].

A number of nanoparticles have been synthesized for use as immunosuppressants for biomedical applications. Such nanoparticles maybe used directly to induce immunosuppression or indirectly when it serves as a drug delivery vehicle [152]. Ref. [154] prepared polylactide-cyclosporine A nanoparticles and found that they induced immunosuppression by suppressing T-cell proliferation. These nanoparticles could also be directed to lymph nodes to induce immunosuppression during drug delivery. In another study, rapamycinloaded chitosan/polylactic acid nanoparticles were reported to induce immunosuppression during corneal transplantation in rabbit models [155]. Carbon nanoparticles and other metal nanoparticles have also been extensively used as immunosuppressants in targeted drug delivery [156–159]. Immunosuppression can also be brought about by tumors which allow them to proliferate in the body. A number of nanoparticles have been synthesized to reverse immunosuppression as a way to destroy tumors. For example, Ref. [160] synthesized self-stabilized hyaluronic acid nanoparticles to deliver chemotherapy drugs to osteosarcoma sites and reported that these nanoparticles reverse immunosuppression brought about by the tumor. A significant decrease in the growth of the tumors was observed as a result of the synergistic effect of immunotherapy and chemotherapy. In another similar study, modified phenylboronic acid nanoparticles inhibited pancreatic tumors and simultaneously alleviated the tumour-induced immunosuppression [161]. Ref. [162] synthesized poly lactic-co-glycolic acid nanoparticles which were coated with PD-L1 (programmed cell death-ligand 1) overexpressed mesenchymal stem cells and induced immunosuppression when administered at a tumor site in the liver of mice. Such nanoparticles have been found to be promising to induce immunosuppression for treatment of tumors and cancers and for effective drug delivery and therapy.

13.5 Summary and conclusions

Nanoparticles are important in a number of biomedical applications. However, nanoparticles such as carbon nanotubes, metallic nanoparticles, and magnetic nanoparticles have been proven to be toxic inducing oxidative stress, DNA damage, decreasing cell viability, and histopathological alterations. These particles can also be translocated to different nontarget organs in the body such as the brain, liver, testes, etc., where they can accumulate and possibly exert toxic effects. The mechanism of toxicity of most nanoparticles is via the generation of free radicals which in turn affect various biomolecules in cells. Therefore, the toxicity of these particles needs to be tested prior to their use. Various biomarkers can be used to test the toxicity of nanomaterials in vivo and in vitro such as the micronucleus test and the comet assay. The use of biocompatible nanoparticles, surface modification, and immunosuppression are some strategies which can be used to minimize potential toxic side effects of nanoparticles. The green synthesis of nanomaterials also appears to minimize toxicity compared to the chemically engineered ones.

13.6 Challenges and future outlook

The progress of nanomaterial research in biomedicine has been profound over the recent years which is evident in successful treatment of a variety of diseases including certain cancers. The availability of variegated form of nanotechnology has fast tracked many medical procedures and treatments which otherwise were time intensive. The rapid blooming of nanoallotropes of different materials gives us great promise in the medical future as drug vectors, as well as being the drugs themselves with their acquired novel chemical and optical properties at the nanoscale level. However enough data regarding the safe and regular usage of certain nanoparticles is still scarce and wanting. Issues with biocompatibility and surface modifications could still potentially arise and studies on long-term effects of these nanoparticles are lacking. Although nanoparticles can be degraded/removed from the body after completing their therapeutic effects, they may still remain accumulated in different tissues and cause toxic side effects. Contradictory findings make it arduous for the researcher and user to build stable formulations for various therapies. The biggest challenge while using NPs is that at different nanoscales, the NP of the same material may exhibit different properties making it difficult to label their action and behavior toward a procedure. The use of animals and cell cultures also may show contrasting results where toxicity of a particular nanoparticle may manifest in either model and not in both.

The use of green chemistry is promising in the field of nanomedicine as nanoparticles are synthesized from natural sources such as plants and are more biocompatible than chemically engineered nanoparticles. Additionally, toxic residues may not be present in green nanoparticles thus making them suitable for use in various biomedical applications. Biocompatibility of nanoparticles should also be tested for all types of toxicity in animal models particularly pertaining to the nervous system and reproductive system in order to fully understand long-term effects.

References

- U. Badıllı, F. Mollarasouli, N.K. Bakirhan, Y. Ozkan, S.A. Ozkan, Role of quantum dots in pharmaceutical and biomedical analysis, and its application in drug delivery, TrAC, Trends Anal. Chem. 131 (2020) 116013, https://doi.org/10.1016/j.trac.2020.116013.
- [2] B. Gidwani, V. Sahu, S.S. Shukla, R. Pandey, V. Joshi, V.K. Jain, et al., Quantum dots: prospectives, toxicity, advances and applications, J. Drug Deliv. Sci. Technol. 61 (2021) 102308, https://doi.org/ 10.1016/j.jddst.2020.102308.
- [3] M.D. Villalva, V. Agarwal, M. Ulanova, P.S. Sachdev, N. Braidy, Quantum dots as a theranostic approach in Alzheimer's disease: a systematic review, Nanomedicine 16 (18) (2021) 1595–1611, https://doi.org/10.2217/nnm-2021-0104.

- [4] V. Amenta, K. Aschberger, Carbon nanotubes: potential medical applications and safety concerns, WIREs Nanomed. Nanobiotechnol. 7 (3) (2015) 371–386, https://doi.org/10.1002/wnan.1317.
- [5] A. Eatemadi, H. Daraee, H. Karimkhanloo, M. Kouhi, N. Zarghami, A. Akbarzadeh, et al., Carbon nanotubes: properties, synthesis, purification, and medical applications, Nanoscale Res. Lett. 9 (1) (2014) 393, https://doi.org/10.1186/1556-276X-9-393.
- [6] H. He, L.A. Pham-Huy, P. Dramou, D. Xiao, P. Zuo, C. Pham-Huy, Carbon nanotubes: applications in pharmacy and medicine, BioMed Res. Int. 2013 (2013) 578290, https://doi.org/10.1155/ 2013/578290.
- [7] H. Daraee, A. Etemadi, M. Kouhi, S. Alimirzalu, A. Akbarzadeh, Application of liposomes in medicine and drug delivery, Artif. Cell Nanomed. Biotechnol. 44 (1) (2016) 381–391, https://doi.org/ 10.3109/21691401.2014.953633.
- [8] E.-M. Kim, H.-J. Jeong, Liposomes: biomedical applications, Chonnam Med. J. 57 (1) (2021) 27–35, https://doi.org/10.4068/cmj.2021.57.1.27.
- N. Tran, T.J. Webster, Magnetic nanoparticles: biomedical applications and challenges, J. Mater. Chem. 20 (40) (2010) 8760-8767, https://doi.org/10.1039/C0JM00994F.
- [10] V.F. Cardoso, A. Francesko, C. Ribeiro, M. Bañobre-López, P. Martins, S. Lanceros-Mendez, Advances in magnetic nanoparticles for biomedical applications, Adv. Healthc. Mater. 7 (5) (2018) 1700845, https://doi.org/10.1002/adhm.201700845.
- [11] H.M. Williams, The application of magnetic nanoparticles in the treatment and monitoring of cancer and infectious diseases, Biosci. Horiz. Int. J. Stud. Res. 10 (2017) hzx009. https://doi.org/10.1093/ biohorizons/hzx009.
- [12] R.E. Bailey, A.M. Smith, S. Nie, Quantum dots in biology and medicine, Phys. E Low-dimens. Syst. Nanostruct. 25 (1) (2004) 1–12, https://doi.org/10.1016/j.physe.2004.07.013.
- [13] J. Lovrić, H.S. Bazzi, Y. Cuie, G.R.A. Fortin, F.M. Winnik, D. Maysinger, Differences in subcellular distribution and toxicity of green and red emitting CdTe quantum dots, J. Mol. Med. 83 (5) (2005) 377–385, https://doi.org/10.1007/s00109-004-0629-x.
- [14] W. Zhang, L. Yang, H. Kuang, P. Yang, Z.P. Aguilar, A. Wang, et al., Acute toxicity of quantum dots on late pregnancy mice: effects of nanoscale size and surface coating, J. Hazard Mater. 318 (2016) 61–69, https://doi.org/10.1016/j.jhazmat.2016.06.048.
- [15] H. Sun, F. Zhang, H. Wei, B. Yang, The effects of composition and surface chemistry on the toxicity of quantum dots, J. Mater. Chem. B 1 (47) (2013) 6485–6494, https://doi.org/10.1039/C3TB21151G.
- [16] M. Wang, J. Wang, H. Sun, S. Han, S. Feng, L. Shi, et al., Time-dependent toxicity of cadmium telluride quantum dots on liver and kidneys in mice: histopathological changes with elevated free cadmium ions and hydroxyl radicals, Int. J. Nanomed. 11 (2016) 2319–2328, https://doi.org/10.2147/ IJN.S103489.
- [17] J.A.J. Fitzpatrick, S.K. Andreko, L.A. Ernst, A.S. Waggoner, B. Ballou, M.P. Bruchez, Long-term persistence and spectral blue shifting of quantum dots in vivo, Nano Lett. 9 (7) (2009) 2736–2741, https://doi.org/10.1021/nl901534q.
- [18] M.M. Haque, H.-Y. Im, J.-E. Seo, M. Hasan, K. Woo, O.-S. Kwon, Acute toxicity and tissue distribution of CdSe/CdS-MPA quantum dots after repeated intraperitoneal injection to mice, J. Appl. Toxicol. 33 (9) (2013) 940–950, https://doi.org/10.1002/jat.2775.
- [19] T. Wu, X. Wang, M. Chen, X. Zhang, J. Zhang, J. Cheng, et al., Respiratory exposure to graphene quantum dots causes fibrotic effects on lung, liver and kidney of mice, Food Chem. Toxicol. 163 (2022) 112971, https://doi.org/10.1016/j.fct.2022.112971.
- [20] F. Ruan, R. Liu, K. Wang, J. Zeng, Z. Zuo, C. He, Y. Zhang, Cytotoxicity of black phosphorus quantum dots on lung-derived cells and the underlying mechanisms, J. Hazard Mater. 402 (2021) 122875, https://doi.org/10.1016/j.jhazmat.2020.122875.
- [21] C.-C. Ho, H. Chang, H.-T. Tsai, M.-H. Tsai, C.-S. Yang, Y.-C. Ling, P. Lin, Quantum dot 705, a cadmium-based nanoparticle, induces persistent inflammation and granuloma formation in the mouse lung, Nanotoxicology 7 (1) (2013) 105–115, https://doi.org/10.3109/17435390.2011.635814.
- [22] J.R. Roberts, J.M. Antonini, D.W. Porter, R.S. Chapman, J.F. Scabilloni, S.-H. Young, et al., Lung toxicity and biodistribution of Cd/Se-ZnS quantum dots with different surface functional groups after pulmonary exposure in rats, Part. Fibre Toxicol. 10 (1) (2013) 5, https://doi.org/10.1186/1743-8977-10-5.

- [23] M.-S. Stan, C. Sima, L.O. Cinteza, A. Dinischiotu, Silicon-based quantum dots induce inflammation in human lung cells and disrupt extracellular matrix homeostasis, FEBS J. 282 (15) (2015) 2914–2929, https://doi.org/10.1111/febs.13330.
- [24] S. Fiorito, A. Serafino, F. Andreola, A. Togna, G. Togna, Toxicity and biocompatibility of carbon nanoparticles, J. Nanosci. Nanotechnol. 6 (3) (2006) 591–599.
- [25] Z. Liu, K. Chen, C. Davis, S. Sherlock, Q. Cao, X. Chen, et al., Drug delivery with carbon nanotubes for in vivo cancer treatment, Cancer Res. 68 (16) (2008) 6652–6660, https://doi.org/10.1158/ 0008-5472.CAN-08-1468.
- [26] P.A. Tran, L. Zhang, T.J. Webster, Carbon nanofibers and carbon nanotubes in regenerative medicine, Adv. Drug Deliv. Rev. 61 (12) (2009) 1097–1114, https://doi.org/10.1016/j.addr.2009.07.010.
- [27] S. Prakash, M. Malhotra, W. Shao, C. Tomaro-Duchesneau, S. Abbasi, Polymeric nanohybrids and functionalized carbon nanotubes as drug delivery carriers for cancer therapy, Adv. Drug Deliv. Rev. 63 (14) (2011) 1340–1351, https://doi.org/10.1016/j.addr.2011.06.013.
- [28] W. Zhang, Z. Zhang, Y. Zhang, The application of carbon nanotubes in target drug delivery systems for cancer therapies, Nanoscale Res. Lett. 6 (1) (2011) 555, https://doi.org/10.1186/1556-276X-6-555.
- [29] K. Soto, K.M. Garza, L.E. Murr, Cytotoxic effects of aggregated nanomaterials, Acta Biomater. 3 (3) (2007) 351–358, https://doi.org/10.1016/j.actbio.2006.11.004.
- [30] Y. Zhu, W. Li, Q. Li, Y. Li, Y. Li, X. Zhang, Q. Huang, Effects of serum proteins on intracellular uptake and cytotoxicity of carbon nanoparticles, Carbon 47 (5) (2009) 1351–1358, https://doi.org/ 10.1016/j.carbon.2009.01.026.
- [31] S. Kang, J.-E. Kim, D. Kim, C.G. Woo, P.V. Pikhitsa, M.-H. Cho, M. Choi, Comparison of cellular toxicity between multi-walled carbon nanotubes and onion-like shell-shaped carbon nanoparticles, J. Nanoparticle Res. 17 (9) (2015) 378, https://doi.org/10.1007/s11051-015-3181-4.
- [32] H.L. Karlsson, P. Cronholm, J. Gustafsson, L. Möller, Copper oxide nanoparticles are highly toxic: a comparison between metal oxide nanoparticles and carbon nanotubes, Chem. Res. Toxicol. 21 (9) (2008) 1726–1732, https://doi.org/10.1021/tx800064j.
- [33] A.A. Shvedova, E.R. Kisin, R. Mercer, A.R. Murray, V.J. Johnson, A.I. Potapovich, et al., Unusual inflammatory and fibrogenic pulmonary responses to single-walled carbon nanotubes in mice, Am. J. Physiol. Lung Cell Mol. Physiol. 289 (5) (2005) L698–L708, https://doi.org/10.1152/ ajplung.00084.2005.
- [34] Y. Liu, Y. Zhao, B. Sun, C. Chen, Understanding the toxicity of carbon nanotubes, Acc. Chem. Res. 46 (3) (2013) 702–713, https://doi.org/10.1021/ar300028m.
- [35] R. Bakry, R.M. Vallant, M. Najam-ul-Haq, M. Rainer, Z. Szabo, C.W. Huck, et al., Medicinal applications of fullerenes, Int. J. Nanomed. 2 (4) (2007) 639–649. https://pubmed.ncbi.nlm.nih.gov/ 18203430.
- [36] S.Z. Mousavi, S. Nafisi, H.I. Maibach, Fullerene nanoparticle in dermatological and cosmetic applications, Nanomed. Nanotechnol. Biol. Med. 13 (3) (2017) 1071–1087, https://doi.org/ 10.1016/j.nano.2016.10.002.
- [37] C.M. Sayes, K.L. Reed, D.B. Warheit, Assessing toxicity of fine and nanoparticles: comparing in vitro measurements to in vivo pulmonary toxicity profiles, Toxicol. Sci. 97 (1) (2007) 163–180, https:// doi.org/10.1093/toxsci/kfm018.
- [38] G.L. Baker, A. Gupta, M.L. Clark, B.R. Valenzuela, L.M. Staska, S.J. Harbo, et al., Inhalation toxicity and lung toxicokinetics of C60 fullerene nanoparticles and microparticles, Toxicol. Sci. 101 (1) (2008) 122–131, https://doi.org/10.1093/toxsci/kfm243.
- [39] B.C. Sayers, D.R. Germolec, N.J. Walker, K.A. Shipkowski, M.D. Stout, M.F. Cesta, et al., Respiratory toxicity and immunotoxicity evaluations of microparticle and nanoparticle C60 fullerene aggregates in mice and rats following nose-only inhalation for 13 weeks, Nanotoxicology 10 (10) (2016) 1458–1468, https://doi.org/10.1080/17435390.2016.1235737.
- [40] N.R. Jacobsen, G. Pojana, P. White, P. Møller, C.A. Cohn, K. Smith Korsholm, et al., Genotoxicity, cytotoxicity, and reactive oxygen species induced by single-walled carbon nanotubes and C60 fullerenes in the FE1-MutaTMMouse lung epithelial cells, Environ. Mol. Mutagen. 49 (6) (2008) 476–487, https://doi.org/10.1002/em.20406.

- [41] Y. Li, K. Song, Y. Cao, C. Peng, G. Yang, Keratin-templated synthesis of metallic oxide nanoparticles as MRI contrast agents and drug carriers, ACS Appl. Mater. Interfaces 10 (31) (2018) 26039–26045, https://doi.org/10.1021/acsami.8b08555.
- [42] F.U. Din, W. Aman, I. Ullah, O.S. Qureshi, O. Mustapha, S. Shafique, A. Zeb, Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors, Int. J. Nanomed. 12 (2017) 7291–7309, https://doi.org/10.2147/IJN.S146315.
- [43] E.R. Evans, P. Bugga, V. Asthana, R. Drezek, Metallic nanoparticles for cancer immunotherapy, Mater. Today 21 (6) (2018) 673–685, https://doi.org/10.1016/j.mattod.2017.11.022.
- [44] M. Wang, X. Lai, L. Shao, L. Li, Evaluation of immunoresponses and cytotoxicity from skin exposure to metallic nanoparticles, Int. J. Nanomed. 13 (2018) 4445–4459, https://doi.org/10.2147/ IJN.S170745.
- [45] S.-D. Li, L. Huang, Nanoparticles evading the reticuloendothelial system: role of the supported bilayer, Biochim. Biophys. Acta 1788 (10) (2009) 2259–2266, https://doi.org/10.1016/ j.bbamem.2009.06.022.
- [46] H.H. Gustafson, D. Holt-Casper, D.W. Grainger, H. Ghandehari, Nanoparticle uptake: the phagocyte problem, Nano Today 10 (4) (2015) 487–510, https://doi.org/10.1016/j.nantod.2015.06.006.
- [47] Y. Tang, X. Wang, J. Li, Y. Nie, G. Liao, Y. Yu, C. Li, Overcoming the reticuloendothelial system barrier to drug delivery with a "don't-eat-us" strategy, ACS Nano 13 (11) (2019) 13015–13026, https://doi.org/10.1021/acsnano.9b05679.
- [48] G. Sonavane, K. Tomoda, K. Makino, Biodistribution of colloidal gold nanoparticles after intravenous administration: effect of particle size, Colloids Surf. B Biointerfaces 66 (2) (2008) 274–280, https://doi.org/10.1016/j.colsurfb.2008.07.004.
- [49] D.P.K. Lankveld, A.G. Oomen, P. Krystek, A. Neigh, A. Troost-de Jong, C.W. Noorlander, et al., The kinetics of the tissue distribution of silver nanoparticles of different sizes, Biomaterials 31 (32) (2010) 8350–8361, https://doi.org/10.1016/j.biomaterials.2010.07.045.
- [50] O.A. Wong, R.J. Hansen, T.W. Ni, C.L. Heinecke, W.S. Compel, D.L. Gustafson, et al., Structure–activity relationships for biodistribution, pharmacokinetics, and excretion of atomically precise nanoclusters in a murine model, Nanoscale 5 (21) (2013) 10525–10533, https://doi.org/ 10.1039/C3NR03121G.
- [51] Y.S. Kim, J.S. Kim, H.S. Cho, D.S. Rha, J.M. Kim, J.D. Park, et al., Twenty-eight-day oral toxicity, genotoxicity, and gender-related tissue distribution of silver nanoparticles in Sprague-Dawley rats, Inhal. Toxicol. 20 (6) (2008) 575–583, https://doi.org/10.1080/08958370701874663.
- [52] M. Baek, H.-E. Chung, J. Yu, J.-A. Lee, T.-H. Kim, J.-M. Oh, et al., Pharmacokinetics, tissue distribution, and excretion of zinc oxide nanoparticles, Int. J. Nanomed. 7 (2012) 3081–3097, https:// doi.org/10.2147/IJN.S32593.
- [53] J.-T. Kwon, A. Minai-Tehrani, S.-K. Hwang, J.-E. Kim, J.-Y. Shin, K.-N. Yu, et al., Acute pulmonary toxicity and body distribution of inhaled metallic silver nanoparticles, Toxicol. Res. 28 (1) (2012) 25–31, https://doi.org/10.5487/TR.2012.28.1.025.
- [54] M. Bednarski, M. Dudek, J. Knutelska, L. Nowiński, J. Sapa, M. Zygmunt, et al., The influence of the route of administration of gold nanoparticles on their tissue distribution and basic biochemical parameters: in vivo studies, Pharmacol. Rep. 67 (3) (2015) 405–409, https://doi.org/10.1016/ j.pharep.2014.10.019.
- [55] X. Zhang, Gold nanoparticles: recent advances in the biomedical applications, Cell Biochem. Biophys. 72 (3) (2015) 771–775, https://doi.org/10.1007/s12013-015-0529-4.
- [56] A.F. Versiani, L.M. Andrade, E.M.N. Martins, S. Scalzo, J.M. Geraldo, C.R. Chaves, et al., Gold nanoparticles and their applications in biomedicine, Future Virol. 11 (4) (2016) 293–309, https:// doi.org/10.2217/fvl-2015-0010.
- [57] K. Sztandera, M. Gorzkiewicz, B. Klajnert-Maculewicz, Gold nanoparticles in cancer treatment, Mol. Pharm. 16 (1) (2019) 1–23, https://doi.org/10.1021/acs.molpharmaceut.8b00810.
- [58] Y.-S. Chen, Y.-C. Hung, I. Liau, G.S. Huang, Assessment of the in vivo toxicity of gold nanoparticles, Nanoscale Res. Lett. 4 (8) (2009) 858, https://doi.org/10.1007/s11671-009-9334-6.
- [59] T. Mironava, M. Hadjiargyrou, M. Simon, V. Jurukovski, M.H. Rafailovich, Gold nanoparticles cellular toxicity and recovery: effect of size, concentration and exposure time, Nanotoxicology 4 (1) (2010) 120–137, https://doi.org/10.3109/17435390903471463.

- [60] N.M. Schaeublin, L.K. Braydich-Stolle, A.M. Schrand, J.M. Miller, J. Hutchison, J.J. Schlager, et al., Surface charge of gold nanoparticles mediates mechanism of toxicity, Nanoscale 3 (2) (2011) 410–420, https://doi.org/10.1039/C0NR00478B.
- [61] A.M. El Badawy, R.G. Silva, B. Morris, K.G. Scheckel, M.T. Suidan, T.M. Tolaymat, Surface charge-dependent toxicity of silver nanoparticles, Environ. Sci. Technol. 45 (1) (2011) 283–287, https://doi.org/10.1021/es1034188.
- [62] T.-H. Kim, M. Kim, H.-S. Park, U.S. Shin, M.-S. Gong, H.-W. Kim, Size-dependent cellular toxicity of silver nanoparticles, J. Biomed. Mater. Res. 100A (4) (2012) 1033–1043, https:// doi.org/10.1002/jbm.a.34053.
- [63] P. Kovvuru, P.E. Mancilla, A.B. Shirode, T.M. Murray, T.J. Begley, R. Reliene, Oral ingestion of silver nanoparticles induces genomic instability and DNA damage in multiple tissues, Nanotoxicology 9 (2) (2015) 162–171, https://doi.org/10.3109/17435390.2014.902520.
- [64] Y.S. Kim, M.Y. Song, J.D. Park, K.S. Song, H.R. Ryu, Y.H. Chung, et al., Subchronic oral toxicity of silver nanoparticles, Part. Fibre Toxicol. 7 (1) (2010) 20, https://doi.org/10.1186/1743-8977-7-20.
- [65] M. van der Zande, R.J. Vandebriel, E. Van Doren, E. Kramer, Z. Herrera Rivera, C.S. Serrano-Rojero, et al., Distribution, elimination, and toxicity of silver nanoparticles and silver ions in rats after 28-day oral exposure, ACS Nano 6 (8) (2012) 7427–7442, https://doi.org/10.1021/nn302649p.
- [66] A. Pratsinis, P. Hervella, J.-C. Leroux, S.E. Pratsinis, G.A. Sotiriou, Toxicity of silver nanoparticles in macrophages, Small 9 (15) (2013) 2576–2584, https://doi.org/10.1002/smll.201202120.
- [67] D. McShan, P.C. Ray, H. Yu, Molecular toxicity mechanism of nanosilver, J. Food Drug Anal. 22 (1) (2014) 116–127, https://doi.org/10.1016/j.jfda.2014.01.010.
- [68] J.S. Teodoro, A.M. Simões, F.V. Duarte, A.P. Rolo, R.C. Murdoch, S.M. Hussain, C.M. Palmeira, Assessment of the toxicity of silver nanoparticles in vitro: a mitochondrial perspective, Toxicol. Vitro 25 (3) (2011) 664–670, https://doi.org/10.1016/j.tiv.2011.01.004.
- [69] J.W. Rasmussen, E. Martinez, P. Louka, D.G. Wingett, Zinc oxide nanoparticles for selective destruction of tumor cells and potential for drug delivery applications, Expert Opin. Drug Deliv. 7 (9) (2010) 1063–1077, https://doi.org/10.1517/17425247.2010.502560.
- [70] W.I. Hagens, A.G. Oomen, W.H. de Jong, F.R. Cassee, A.J.A.M. Sips, What do we (need to) know about the kinetic properties of nanoparticles in the body? Regul. Toxicol. Pharmacol. 49 (3) (2007) 217–229, https://doi.org/10.1016/j.yrtph.2007.07.006.
- [71] S. Lanone, J. Boczkowski, Biomedical applications and potential health risks of nanomaterials: molecular mechanisms, Curr. Mol. Med. 6 (6) (2006) 651–663, https://doi.org/10.2174/ 156652406778195026.
- [72] W.-S. Cho, R. Duffin, S.E.M. Howie, C.J. Scotton, W.A.H. Wallace, W. MacNee, et al., Progressive severe lung injury by zinc oxide nanoparticles; the role of Zn²⁺ dissolution inside lysosomes, Part. Fibre Toxicol. 8 (1) (2011) 27, https://doi.org/10.1186/1743-8977-8-27.
- [73] N. Andre, X. Tian, M. Lutz, L. Ning, Toxic potential of materials at the nanolevel, Science 311 (5761) (2006) 622–627, https://doi.org/10.1126/science.1114397.
- [74] H.A. Jeng, J. Swanson, Toxicity of metal oxide nanoparticles in mammalian cells, J. Environ. Sci. Health Part A 41 (12) (2006) 2699–2711, https://doi.org/10.1080/10934520600966177.
- [75] T. Xia, M. Kovochich, J. Brant, M. Hotze, J. Sempf, T. Oberley, et al., Comparison of the abilities of ambient and manufactured nanoparticles to induce cellular toxicity according to an oxidative stress paradigm, Nano Lett. 6 (2006) 1794–1807, https://doi.org/10.1021/nl061025k.
- [76] W. Lin, Y. Xu, C.-C. Huang, Y. Ma, K. Shannon, D.-R. Chen, Toxicity of nano- and micro-sized ZnO particles in human lung epithelial cells, J. Nanoparticle Res. 11 (2008) 25–39, https://doi.org/ 10.1007/s11051-008-9419-7.
- [77] C.M. Sayes, A.A. Marchione, K.L. Reed, D.B. Warheit, Comparative pulmonary toxicity assessments of C60 water suspensions in rats: few differences in fullerene toxicity in vivo in contrast to in vitro profiles, Nano Lett. 7 (8) (2007) 2399–2406, https://doi.org/10.1021/nl0710710.
- [78] J. Sanchez-Betancourt, V. Anaya-Martínez, A.L. Gutierrez-Valdez, J.L. Ordoñez-Librado, E. Montiel-Flores, J. Espinosa-Villanueva, et al., Manganese mixture inhalation is a reliable Parkinson disease model in rats, Neurotoxicology 33 (5) (2012) 1346–1355, https://doi.org/10.1016/ j.neuro.2012.08.012.

- [79] J. Bornhorst, S. Meyer, T. Weber, C. Böker, T. Marschall, A. Mangerich, et al., Molecular mechanisms of Mn induced neurotoxicity: RONS generation, genotoxicity, and DNA-damage response, Mol. Nutr. Food Res. 57 (7) (2013) 1255–1269, https://doi.org/10.1002/mnfr.201200758.
- [80] T. Verina, J.S. Schneider, T.R. Guilarte, Manganese exposure induces α-synuclein aggregation in the frontal cortex of non-human primates, Toxicol. Lett. 217 (3) (2013) 177–183, https://doi.org/ 10.1016/j.toxlet.2012.12.006.
- [81] W.W. Dlamini, G. Nelson, S.S. Nielsen, B.A. Racette, Manganese exposure, parkinsonian signs, and quality of life in South African mine workers, Am. J. Ind. Med. 63 (1) (2020) 36–43, https://doi.org/ 10.1002/ajim.23060.
- [82] S.M. Hussain, A.K. Javorina, A.M. Schrand, H.M. Duhart, S.F. Ali, J.J. Schlager, The interaction of manganese nanoparticles with PC-12 cells induces dopamine depletion, Toxicol. Sci. 92 (2) (2006) 456–463, https://doi.org/10.1093/toxsci/kfl020.
- [83] N. Yousefalizadegan, Z. Mousavi, T. Rastegar, Y. Razavi, P. Najafizadeh, Reproductive toxicity of manganese dioxide in forms of micro- and nanoparticles in male rats, Int. J. Reprod. Biomed. 17 (5) (2018) 361–370, https://doi.org/10.18502/ijrm.v17i5.4603.
- [84] S.P. Singh, M. Kumari, S.I. Kumari, M.F. Rahman, M. Mahboob, P. Grover, Toxicity assessment of manganese oxide micro and nanoparticles in Wistar rats after 28 days of repeated oral exposure, J. Appl. Toxicol. 33 (10) (2013) 1165–1179, https://doi.org/10.1002/jat.2887.
- [85] T. Guo, M. Lin, J. Huang, C. Zhou, W. Tian, H. Yu, et al., The recent advances of magnetic nanoparticles in medicine, J. Nanomater. 2018 (2018) 7805147, https://doi.org/10.1155/2018/7805147.
- [86] J.S. Kim, T.-J. Yoon, K.N. Yu, B.G. Kim, S.J. Park, H.W. Kim, et al., Toxicity and tissue distribution of magnetic nanoparticles in mice, Toxicol. Sci. 89 (1) (2006) 338–347, https://doi.org/ 10.1093/toxsci/kfj027.
- [87] U.O. Häfeli, J.S. Riffle, L. Harris-Shekhawat, A. Carmichael-Baranauskas, F. Mark, J.P. Dailey, D. Bardenstein, Cell uptake and in vitro toxicity of magnetic nanoparticles suitable for drug delivery, Mol. Pharm. 6 (5) (2009) 1417–1428, https://doi.org/10.1021/mp900083m.
- [88] R.M. Amin, A. Abdelmonem, T. Verwanger, E. Elsherbini, B. Krammer, Cytotoxicity of magnetic nanoparticles on normal and malignant human skin cells, Nano Life 04 (01) (2013) 1440002, https:// doi.org/10.1142/S1793984414400029.
- [89] R. Mejías, L. Gutiérrez, G. Salas, S. Pérez-Yagüe, T.M. Zotes, F.J. Lázaro, et al., Long term biotransformation and toxicity of dimercaptosuccinic acid-coated magnetic nanoparticles support their use in biomedical applications, J. Control. Release 171 (2) (2013) 225–233, https://doi.org/10.1016/ j.jconrel.2013.07.019.
- [90] A.M. Prodan, S.L. Iconaru, C.S. Ciobanu, M.C. Chifiriuc, M. Stoicea, D. Predoi, Iron oxide magnetic nanoparticles: characterization and toxicity evaluation by in vitro and in vivo assays, J. Nanomater. 2013 (2013), https://doi.org/10.1155/2013/587021.
- [91] Y.G. Toropova, A.S. Golovkin, A.B. Malashicheva, D.V. Korolev, A.N. Gorshkov, K.G. Gareev, et al., In vitro toxicity of FemOn, FemOn-SiO2 composite, and SiO2-FemOn core-shell magnetic nanoparticles, Int. J. Nanomed. 12 (2017) 593–603, https://doi.org/10.2147/IJN.S122580.
- [92] A. Akbarzadeh, M. Samiei, S. Davaran, Magnetic nanoparticles: preparation, physical properties, and applications in biomedicine, Nanoscale Res. Lett. 7 (1) (2012) 144, https://doi.org/10.1186/1556-276X-7-144.
- [93] O. Baber, M. Jang, D. Barber, K. Powers, Amorphous silica coatings on magnetic nanoparticles enhance stability and reduce toxicity to in vitro BEAS-2B cells, Inhal. Toxicol. 23 (9) (2011) 532-543, https://doi.org/10.3109/08958378.2011.592869.
- [94] J.-T. Kwon, S.-K. Hwang, H. Jin, D.-S. Kim, A. Minai-Tehrani, H.-J. Yoon, et al., Body distribution of inhaled fluorescent magnetic nanoparticles in the mice, J. Occup. Health 50 (1) (2008) 1–6, https://doi.org/10.1539/joh.50.1.
- [95] J.-T. Kwon, D.-S. Kim, A. Minai-Tehrani, S.-K. Hwang, S.-H. Chang, E.-S. Lee, et al., Inhaled fluorescent magnetic nanoparticles induced extramedullary hematopoiesis in the spleen of mice, J. Occup. Health (2009) advpub, 908110093, https://doi.org/10.1539/joh.L8159.

- [96] A.H. D'Costa, S.K. Shyama, P.K. MK, V.S. Verenkar, R.B. Tangsali, Genotoxic effect of manganese and nickel doped zinc ferrite (Mn 0.3 Ni 0.3 Zn 0.4 Fe 2 O 4) nanoparticle in Swiss albino mouse *Mus musculus*, Indian J. Exp. Biol. 59 (01) (2020) 25–32.
- [97] L. Sadeghi, V. Yousefi Babadi, H.R. Espanani, Toxic effects of the Fe₂O₃ nanoparticles on the liver and lung tissue, Bratisl. Lek. Listy 116 (6) (2015) 373–378, https://doi.org/10.4149/bll_2015_071.
- [98] A. Nemmar, S. Beegam, P. Yuvaraju, J. Yasin, S. Tariq, S. Attoub, B.H. Ali, Ultrasmall superparamagnetic iron oxide nanoparticles acutely promote thrombosis and cardiac oxidative stress and DNA damage in mice, Part. Fibre Toxicol. 13 (1) (2016) 22, https://doi.org/10.1186/s12989-016-0132-x.
- [99] U.A. Reddy, P.V. Prabhakar, M. Mahboob, Biomarkers of oxidative stress for in vivo assessment of toxicological effects of iron oxide nanoparticles, Saudi J. Biol. Sci. 24 (6) (2017) 1172–1180, https:// doi.org/10.1016/j.sjbs.2015.09.029.
- [100] T. Mai, J.Z. Hilt, Magnetic nanoparticles: reactive oxygen species generation and potential therapeutic applications, J. Nanoparticle Res. 19 (7) (2017) 253, https://doi.org/10.1007/s11051-017-3943-2.
- [101] X.-Y. Lu, D.-C. Wu, Z.-J. Li, G.-Q. Chen, Chpater 7—polymer nanoparticles, in: A. B. T.-P. in M. B, T.S. Villaverde (Eds.), Nanoparticles in Translational Science and Medicine vol. 104, Academic Press, 2011, pp. 299–323, https://doi.org/10.1016/B978-0-12-416020-0.00007-3.
- [102] R. Shah, D. Eldridge, E. Palombo, I. Harding, in: Introduction BT—Lipid Nanoparticles: Production, Characterization and Stability, Springer International Publishing, 2015, pp. 1–9, https:// doi.org/10.1007/978-3-319-10711-0_1.
- [103] N. Voigt, P. Henrich-Noack, S. Kockentiedt, W. Hintz, J. Tomas, B.A. Sabel, Toxicity of polymeric nanoparticles in vivo and in vitro, J. Nanoparticle Res. 16 (6) (2014) 2379, https://doi.org/10.1007/ s11051-014-2379-1.
- [104] N. Grabowski, H. Hillaireau, J. Vergnaud, N. Tsapis, M. Pallardy, S. Kerdine-Römer, et al., Surface coating mediates the toxicity of polymeric nanoparticles towards human-like macrophages, Int. J. Pharm. 482 (1) (2015) 75–83, https://doi.org/10.1016/j.ijpharm.2014.11.042.
- [105] R.P. Singh, P. Ramarao, Accumulated polymer degradation products as effector molecules in cytotoxicity of polymeric nanoparticles, Toxicol. Sci. 136 (1) (2013) 131–143, https://doi.org/10.1093/ toxsci/kft179.
- [106] Y. Panahi, M. Farshbaf, M. Mohammadhosseini, M. Mirahadi, R. Khalilov, S. Saghfi, A. Akbarzadeh, Recent advances on liposomal nanoparticles: synthesis, characterization and biomedical applications, Artif. Cell Nanomed. Biotechnol. 45 (4) (2017) 788–799, https://doi.org/ 10.1080/21691401.2017.1282496.
- [107] P. Milla, F. Dosio, L. Cattel, PEGylation of proteins and liposomes: a powerful and flexible strategy to improve the drug delivery, Curr. Drug Metabol. 13 (1) (2012) 105–119.
- [108] K.B. Knudsen, H. Northeved, P. Kumar EK, A. Permin, T. Gjetting, T.L. Andresen, et al., In vivo toxicity of cationic micelles and liposomes, Nanomed. Nanotechnol. Biol. Med. 11 (2) (2015) 467–477, https://doi.org/10.1016/j.nano.2014.08.004.
- [109] M. Roursgaard, K.B. Knudsen, H. Northeved, M. Persson, T. Christensen, P.E.K. Kumar, et al., In vitro toxicity of cationic micelles and liposomes in cultured human hepatocyte (HepG2) and lung epithelial (A549) cell lines, Toxicol. Vitro 36 (2016) 164–171, https://doi.org/10.1016/ j.tiv.2016.08.002.
- [110] L.M. Kaminskas, V.M. McLeod, B.D. Kelly, G. Sberna, B.J. Boyd, M. Williamson, et al., A comparison of changes to doxorubicin pharmacokinetics, antitumor activity, and toxicity mediated by PEGylated dendrimer and PEGylated liposome drug delivery systems, Nanomed. Nanotechnol. Biol. Med. 8 (1) (2012) 103–111, https://doi.org/10.1016/j.nano.2011.05.013.
- [111] É. de Melo Reis, A.A.A. de Rezende, D.V. Santos, P.F. de Oliveria, H.D. Nicolella, D.C. Tavares, et al., Assessment of the genotoxic potential of two zinc oxide sources (amorphous and nanoparticles) using the in vitro micronucleus test and the in vivo wing somatic mutation and recombination test, Food Chem. Toxicol. 84 (2015) 55–63, https://doi.org/10.1016/j.fct.2015.07.008.
- [112] S. Könen-Adıgüzel, S. Ergene, In vitro evaluation of the genotoxicity of CeO₂ nanoparticles in human peripheral blood lymphocytes using cytokinesis-block micronucleus test, comet assay, and gamma H2AX, Toxicol. Ind. Health 34 (5) (2018) 293–300, https://doi.org/10.1177/ 0748233717753780.

- [113] Y. Li, D.H. Chen, J. Yan, Y. Chen, R.A. Mittelstaedt, Y. Zhang, et al., Genotoxicity of silver nanoparticles evaluated using the Ames test and in vitro micronucleus assay, Mutat. Res. Genet. Toxicol. Environ. Mutagen. 745 (1) (2012) 4–10, https://doi.org/10.1016/j.mrgentox.2011.11.010.
- [114] J. Wang, B. Che, L.W. Zhang, G. Dong, Q. Luo, L. Xin, Comparative genotoxicity of silver nanoparticles in human liver HepG2 and lung epithelial A549 cells, J. Appl. Toxicol. 37 (4) (2017) 495–501, https://doi.org/10.1002/jat.3385.
- [115] J.S. Kim, J.H. Sung, J.H. Ji, K.S. Song, J.H. Lee, C.S. Kang, I.J. Yu, In vivo genotoxicity of silver nanoparticles after 90-day silver nanoparticle inhalation exposure, Saf. Health Work 2 (1) (2011) 34–38, https://doi.org/10.5491/SHAW.2011.2.1.34.
- [116] A.H. D'Costa, S.K. Shyama, M.K. PraveenKumar, V.M.S. Verenkar, R.B. Tangsali, Genotoxic Effect of Manganese and Nickel Doped Zinc Ferrite (Mn Sub (0.3) Ni Sub (0.3) Zn Sub (0.4) Fe Sub (2) O Sub (4)) Nanoparticle in Swiss Albino Mouse Mus musculus, 2021.
- [117] Z. Magdolenova, A. Collins, A. Kumar, A. Dhawan, V. Stone, M. Dusinska, Mechanisms of genotoxicity. A review of in vitro and in vivo studies with engineered nanoparticles, Nanotoxicology 8 (3) (2014) 233–278, https://doi.org/10.3109/17435390.2013.773464.
- [118] L. Song, M.G. Vijver, W.J.G.M. Peijnenburg, T.S. Galloway, C.R. Tyler, A comparative analysis on the in vivo toxicity of copper nanoparticles in three species of freshwater fish, Chemosphere 139 (2015) 181–189, https://doi.org/10.1016/j.chemosphere.2015.06.021.
- [119] Y. Li, T. Qin, T. Ingle, J. Yan, W. He, J.-J. Yin, T. Chen, Differential genotoxicity mechanisms of silver nanoparticles and silver ions, Arch. Toxicol. 91 (1) (2017) 509–519, https://doi.org/10.1007/ s00204-016-1730-y.
- [120] S. Vandghanooni, M. Eskandani, Comet assay: a method to evaluate genotoxicity of nano-drug delivery system, Bioimpacts BI 1 (2) (2011) 87–97, https://doi.org/10.5681/bi.2011.012.
- [121] J. Kain, H.L. Karlsson, L. Möller, DNA damage induced by micro- and nanoparticles—interaction with FPG influences the detection of DNA oxidation in the comet assay, Mutagenesis 27 (4) (2012) 491–500, https://doi.org/10.1093/mutage/ges010.
- [122] M. Naya, N. Kobayashi, M. Ema, S. Kasamoto, M. Fukumuro, S. Takami, et al., In vivo genotoxicity study of titanium dioxide nanoparticles using comet assay following intratracheal instillation in rats, Regul. Toxicol. Pharmacol. 62 (1) (2012) 1–6, https://doi.org/10.1016/j.yrtph.2011.12.002.
- [123] S. Patel, P. Patel, S.R. Bakshi, Titanium dioxide nanoparticles: an in vitro study of DNA binding, chromosome aberration assay, and comet assay, Cytotechnology 69 (2) (2017) 245–263, https:// doi.org/10.1007/s10616-016-0054-3.
- [124] V. Bastos, I.F. Duarte, C. Santos, H. Oliveira, Genotoxicity of citrate-coated silver nanoparticles to human keratinocytes assessed by the comet assay and cytokinesis blocked micronucleus assay, Environ. Sci. Pollut. Control Ser. 24 (5) (2017) 5039–5048, https://doi.org/10.1007/s11356-016-8240-6.
- [125] S. Naahidi, M. Jafari, F. Edalat, K. Raymond, A. Khademhosseini, P. Chen, Biocompatibility of engineered nanoparticles for drug delivery, J. Control. Release 166 (2) (2013) 182–194, https:// doi.org/10.1016/j.jconrel.2012.12.013.
- [126] M.A. Malvindi, V. De Matteis, A. Galeone, V. Brunetti, G.C. Anyfantis, A. Athanassiou, et al., Toxicity assessment of silica coated iron oxide nanoparticles and biocompatibility Improvement by surface engineering, PLoS One 9 (1) (2014) e85835, https://doi.org/10.1371/ journal.pone.0085835.
- [127] H. Nosrati, M. Salehiabar, E. Attari, S. Davaran, H. Danafar, H.K. Manjili, Green and one-pot surface coating of iron oxide magnetic nanoparticles with natural amino acids and biocompatibility investigation, Appl. Organomet. Chem. 32 (2) (2018) e4069, https://doi.org/10.1002/aoc.4069.
- [128] M. Ayubi, M. Karimi, S. Abdpour, K. Rostamizadeh, M. Parsa, M. Zamani, et al., Magnetic nanoparticles decorated with PEGylated curcumin as dual targeted drug delivery: synthesis, toxicity and biocompatibility study, Mater. Sci. Eng. C 104 (2019) 109810, https://doi.org/10.1016/ j.msec.2019.109810.
- [129] G.S. Zamay, T.N. Zamay, K.A. Lukyanenko, A.S. Kichkailo, Aptamers increase biocompatibility and reduce the toxicity of magnetic nanoparticles used in biomedicine, Biomedicines 8 (3) (2020), https://doi.org/10.3390/biomedicines8030059.

- [130] E. Rozhina, A. Danilushkina, F. Akhatova, R. Fakhrullin, A. Rozhin, S. Batasheva, Biocompatibility of magnetic nanoparticles coating with polycations using A549 cells, J. Biotechnol. 325 (2021) 25–34, https://doi.org/10.1016/j.jbiotec.2020.12.003.
- [131] D. Pooja, S. Panyaram, H. Kulhari, S.S. Rachamalla, R. Sistla, Xanthan gum stabilized gold nanoparticles: characterization, biocompatibility, stability and cytotoxicity, Carbohydr. Polym. 110 (2014) 1–9, https://doi.org/10.1016/j.carbpol.2014.03.041.
- [132] E.I. Alarcon, K. Udekwu, M. Skog, N.L. Pacioni, K.G. Stamplecoskie, M. González-Béjar, et al., The biocompatibility and antibacterial properties of collagen-stabilized, photochemically prepared silver nanoparticles, Biomaterials 33 (19) (2012) 4947–4956, https://doi.org/10.1016/ j.biomaterials.2012.03.033.
- [133] L. Grenho, C.L. Salgado, M.H. Fernandes, F.J. Monteiro, M.P. Ferraz, Antibacterial activity and biocompatibility of three-dimensional nanostructured porous granules of hydroxyapatite and zinc oxide nanoparticles—an *in vitro* and *in vivo* study, Nanotechnology 26 (31) (2015) 315101, https://doi.org/10.1088/0957-4484/26/31/315101.
- [134] A.N. Emam, S.A. Loutfy, A.A. Mostafa, H. Awad, M.B. Mohamed, Cyto-toxicity, biocompatibility and cellular response of carbon dots—plasmonic based nano-hybrids for bioimaging, RSC Adv. 7 (38) (2017) 23502–23514, https://doi.org/10.1039/C7RA01423F.
- [135] X. Zhu, T. Zhao, Z. Nie, Z. Miao, Y. Liu, S. Yao, Nitrogen-doped carbon nanoparticle modulated turn-on fluorescent probes for histidine detection and its imaging in living cells, Nanoscale 8 (4) (2016) 2205–2211, https://doi.org/10.1039/C5NR07826A.
- [136] X. Xu, K. Zhang, L. Zhao, C. Li, W. Bu, Y. Shen, et al., Aspirin-based carbon dots, a good biocompatibility of material applied for bioimaging and anti-inflammation, ACS Appl. Mater. Interfaces 8 (48) (2016) 32706–32716, https://doi.org/10.1021/acsami.6b12252.
- [137] A. Gupta, T.A. Liberati, S.J. Verhulst, B.J. Main, M.H. Roberts, A.G.R. Potty, et al., Biocompatibility of single-walled carbon nanotube composites for bone regeneration, Bone Jt. Res. 4 (5) (2015) 70–77, https://doi.org/10.1302/2046-3758.45.2000382.
- [138] S. Iravani, Green synthesis of metal nanoparticles using plants, Green Chem. 13 (10) (2011) 2638–2650, https://doi.org/10.1039/C1GC15386B.
- [139] I. Hussain, N.B. Singh, A. Singh, H. Singh, S.C. Singh, Green synthesis of nanoparticles and its potential application, Biotechnol. Lett. 38 (4) (2016) 545–560, https://doi.org/10.1007/s10529-015-2026-7.
- [140] A. Rana, K. Yadav, S. Jagadevan, A comprehensive review on green synthesis of nature-inspired metal nanoparticles: mechanism, application and toxicity, J. Clean. Prod. 272 (2020) 122880, https://doi.org/10.1016/j.jclepro.2020.122880.
- [141] R. Amooaghaie, M.R. Saeri, M. Azizi, Synthesis, characterization and biocompatibility of silver nanoparticles synthesized from Nigella sativa leaf extract in comparison with chemical silver nanoparticles, Ecotoxicol. Environ. Saf. 120 (2015) 400–408, https://doi.org/10.1016/ j.ecoenv.2015.06.025.
- [142] A.K. Mittal, U.C. Banerjee, In vivo safety, toxicity, biocompatibility and anti-tumour efficacy of bioinspired silver and selenium nanoparticles, Mater. Today Commun. 26 (2021) 102001, https:// doi.org/10.1016/j.mtcomm.2020.102001.
- [143] S.K. Nune, N. Chanda, R. Shukla, K. Katti, R.R. Kulkarni, S. Thilakavathy, et al., Green nanotechnology from tea: phytochemicals in tea as building blocks for production of biocompatible gold nanoparticles, J. Mater. Chem. 19 (19) (2009) 2912–2920, https://doi.org/10.1039/B822015H.
- [144] S. Salatin, S. Maleki Dizaj, A. Yari Khosroushahi, Effect of the surface modification, size, and shape on cellular uptake of nanoparticles, Cell Biol. Int. 39 (8) (2015) 881–890, https://doi.org/10.1002/ cbin.10459.
- [145] X. Chu, J. Yu, Y.-L. Hou, Surface modification of magnetic nanoparticles in biomedicine, Chin. Phys. B 24 (1) (2015) 14704, https://doi.org/10.1088/1674-1056/24/1/014704.
- [146] Y.-K. Chang, Y.-P. Liu, J.H. Ho, S.-C. Hsu, O.K. Lee, Amine-surface-modified superparamagnetic iron oxide nanoparticles interfere with differentiation of human mesenchymal stem cells, J. Orthop. Res. 30 (9) (2012) 1499–1506, https://doi.org/10.1002/jor.22088.

- [147] S. Hong, K.Y. Kim, H.J. Wook, S.Y. Park, K.D. Lee, D.Y. Lee, et al., Attenuation of the in vivo toxicity of biomaterials by polydopamine surface modification, Nanomedicine 6 (5) (2011) 793-801, https://doi.org/10.2217/nnm.11.76.
- [148] S. Manoochehri, B. Darvishi, G. Kamalinia, M. Amini, M. Fallah, S.N. Ostad, et al., Surface modification of PLGA nanoparticles via human serum albumin conjugation for controlled delivery of docetaxel, Daru 21 (1) (2013) 58, https://doi.org/10.1186/2008-2231-21-58.
- [149] B.Q. Wang, P.H. Zhang, M.H. Fang, Potassium dichromate surface modification of poly-lactic acid (PLA) and poly-lactide-co-glycolide (PGLA) fibers, Adv. Mater. Res. 936 (2014) 801–805, https:// doi.org/10.4028/www.scientific.net/AMR.936.801.
- [150] A. Scislowska-Czarnecka, D. Szmigiel, M. Genet, C. Dupont-Gillain, E. Pamula, E. Kolaczkowska, Oxygen plasma surface modification augments poly(L-lactide-co-glycolide) cytocompatibility toward osteoblasts and minimizes immune activation of macrophages, J. Biomed. Mater. Res. 103 (12) (2015) 3965–3977, https://doi.org/10.1002/jbm.a.35509.
- [151] B.S. Zolnik, A. González-Fernández, N. Sadrieh, M.A. Dobrovolskaia, Nanoparticles and the immune system, Endocrinology 151 (2) (2010) 458–465, https://doi.org/10.1210/en.2009-1082.
- [152] T.A. Ngobili, M.A. Daniele, Nanoparticles and direct immunosuppression, Exp. Biol. Med. (Maywood, N.J.) 241 (10) (2016) 1064–1073, https://doi.org/10.1177/1535370216650053.
- [153] P. Ray, N. Haideri, I. Haque, O. Mohammed, S. Chakraborty, S. Banerjee, et al., The impact of nanoparticles on the immune system: a gray zone of nanomedicine, J. Immunol. Sci. 5 (1) (2021).
- [154] J. Azzi, L. Tang, R. Moore, R. Tong, N. El Haddad, T. Akiyoshi, et al., Polylactide-cyclosporin A nanoparticles for targeted immunosuppression, Faseb. J. 24 (10) (2010) 3927–3938, https://doi.org/ 10.1096/fj.10-154690.
- [155] X.-B. Yuan, Y.-B. Yuan, W. Jiang, J. Liu, E.-J. Tian, H.-M. Shun, et al., Preparation of rapamycinloaded chitosan/PLA nanoparticles for immunosuppression in corneal transplantation, Int. J. Pharm. 349 (1) (2008) 241–248, https://doi.org/10.1016/j.ijpharm.2007.07.045.
- [156] R. Huq, E.L.G. Samuel, W.K.A. Sikkema, L.G. Nilewski, T. Lee, M.R. Tanner, et al., Preferential uptake of antioxidant carbon nanoparticles by T lymphocytes for immunomodulation, Sci. Rep. 6 (1) (2016) 33808, https://doi.org/10.1038/srep33808.
- [157] J. Hwang, E. Lee, J. Kim, Y. Seo, K.H. Lee, J.W. Hong, et al., Effective delivery of immunosuppressive drug molecules by silica coated iron oxide nanoparticles, Colloids Surf. B Biointerfaces 142 (2016) 290–296, https://doi.org/10.1016/j.colsurfb.2016.01.040.
- [158] I.-C. Lee, J.-W. Ko, S.-H. Park, N.-R. Shin, I.-S. Shin, C. Moon, et al., Copper nanoparticles induce early fibrotic changes in the liver via TGF-β/Smad signaling and cause immunosuppressive effects in rats, Nanotoxicology 12 (6) (2018) 637–651, https://doi.org/10.1080/17435390.2018.1472313.
- [159] L. Li, M. Zhen, H. Wang, Z. Sun, W. Jia, Z. Zhao, et al., Functional gadofullerene nanoparticles trigger robust cancer immunotherapy based on rebuilding an immunosuppressive tumor microenvironment, Nano Lett. 20 (6) (2020) 4487–4496, https://doi.org/10.1021/acs.nanolett.0c01287.
- [160] Y. Zhang, T. Yuan, Z. Li, C. Luo, Y. Wu, J. Zhang, et al., Hyaluronate-based self-stabilized nanoparticles for immunosuppression reversion and immunochemotherapy in osteosarcoma treatment, ACS Biomater. Sci. Eng. 7 (4) (2021) 1515–1525, https://doi.org/10.1021/acsbiomaterials.1c00081.
- [161] Z. Lu, Y. Long, Y. Wang, X. Wang, C. Xia, M. Li, et al., Phenylboronic acid modified nanoparticles simultaneously target pancreatic cancer and its metastasis and alleviate immunosuppression, Eur. J. Pharm. Biopharm. 165 (2021) 164–173, https://doi.org/10.1016/j.ejpb.2021.05.014.
- [162] S. Shen, H. Dai, Z. Fei, Y. Chai, Y. Hao, Q. Fan, et al., Immunosuppressive nanoparticles for management of immune-related adverse events in liver, ACS Nano 15 (5) (2021) 9111–9125, https:// doi.org/10.1021/acsnano.1c02391.